

# ***VIDYASAGAR UNIVERSITY***



***MIDNAPORE -721 102, WEST BENGAL, INDIA***

Faculty of Science  
**Department of Zoology**

***The Syllabus for POST- GRADUATE Programme  
in  
Zoology***

Under Choice Based Credit System (CBCS)

[w.e.f. : 2018-19]

## **PROGRAMME OUTCOME**

*The course curriculum of the department has been designed and developed giving due importance to both classical components of subject Zoology along with its different ramifications in tune with the modern scientific development across the world. The classical aspects of zoology includes basic structural functional organization of the Non-Chordates and Chordates, taxonomy, biosystematics, histology, animal physiology, evolution and adaptation whereas the modern dimension of the course curriculum has attached most significant aspects of molecular biology, genetics, biotechnology, bioinformatics, biochemistry, immunology and biostatistics. The effort appears to be very fruitful by virtue of the results and academic performance of the existing and former students which are being reflected in their success in different national and state level competition. Keeping pace with the above mentioned course curriculum the department although has initiated its journey with only one special paper now it is offering four different special papers (Fishery, Ecology, Genetics & Molecular biology and parasitology) which also enabled rural based students of the University to get proper recognition both nationally and also getting different subject based services. Moreover the outreach and extension program involving the students in their practical curriculum have provided significant contribution of livelihood generation of marginalized people of the region. All the courses in the program are carefully designed to equip the students for competitive exams like CSIR NET, SET etc. and also to write research proposals for grants.*

## COURSE STRUCTURE OF M.Sc. IN ZOOLOGY

SEM-ESTER	PAPER CODE	CONTENT	Marks	Credit	
I	ZOO 101	Non- Chordates & Chordates			
		ZOO 101.1	Non- Chordates	25	2
		ZOO 101.2	Chordates	25	2
	ZOO 102	Histochemistry& Animal Physiology			
		ZOO 102.1	Histochemistry	25	2
		ZOO 102.2	Animal Physiology	25	2
	ZOO 103	Immunology and Methods in Biology			
		ZOO 103.1	Immunology	25	2
		ZOO 103.2	Methods in Biology	25	2
	ZOO 104	Cell Biology &Cytogenetics			
		ZOO 104.1	Cell Biology	25	2
		ZOO 104.2	Cytogenetics	25	2
	ZOO 195	Non- Chordates, Chordates, Histochemistry& Animal Physiology (Practical)		50	4
ZOO 196	Immunology, Methods in Biology, Cell Biology & Cytogenetics (Practical)		50	4	
<b>TOTAL</b>			<b>300</b>	<b>24</b>	
II	ZOO 201	Biosystematics & Ecological Principles			
		ZOO 201.1	Biosystematics	25	2
		ZOO 201.2	Ecological Principles	25	2
	ZOO 202	Biophysics &Biochemistry			
		ZOO 202.1	Biophysics	25	2
		ZOO 202.2	Biochemistry	25	2
	ZOO 203	Molecular Biology &Parasitology			
		ZOO 203.1	Molecular Biology	25	2
		ZOO 203.2	Parasitology	25	2
	<b>C-ZOO 204</b>	<b>Wildlife &amp; Eco-Management and Aqua informatics</b>			
		C-ZOO 204.1	<b>Wildlife &amp; Eco-Management</b>	25	2
		C-ZOO 204.2	<b>Aqua informatics</b>	25	2
	ZOO 295	Ecological principles, Biochemistry & Field Study		50	4
ZOO 296	Biosystematics, Molecular Biology, Parasitology & Biophysics		50	4	
<b>TOTAL</b>			<b>300</b>	<b>24</b>	
III	ZOO 301	Basic & Applied Entomology and Ecotoxicology			
		ZOO 301.1	Basic & Applied Entomology	25	2
		ZOO 301.2	Ecotoxicology	25	2
	ZOO 302	Molecular Evolution and Microbiology			
		ZOO 302.1	Molecular Evolution	25	2
	ZOO 302.2	Microbiology	25	2	

SPECIAL PAPER (A:Fishery; B:Ecology; C:Genetics & Mol. Biology; D:Prasitology)(one paper)					
ZOO 303A	Fish Taxonomy & Biology and Oceanography				
	ZOO 303A.1	Fish Taxonomy & Biology	25	2	
	ZOO 303A.2	Oceanography	25	2	
ZOO 303B	Biodiversity and Conservation Ecology & Aquatic Ecology				
	ZOO 303B.1	Biodiversity and Conservation Ecology	25	2	
	ZOO 303B.2	Aquatic Ecology	25	2	
ZOO 303C	Genetics & Molecular Biology				
	ZOO 303C.1	Genetics	25	2	
	ZOO 303C.2	Molecular Biology	25	2	
ZOO 303D	Diversity & Biology of Parasite and Immunoparasitology				
	ZOO 303D.1	Diversity and Biology of Parasite	25	2	
	ZOO 303D.2	Immunoparasitology	25	2	
C-ZOO 304	<b>Genetics and Haematology(CBCS)</b>				
	C-ZOO 304.1	<b>Genetics</b>	25	2	
	C-ZOO 304.2	<b>Haematology</b>	25	2	
ZOO 395	Entomology, Ecotoxicology, Molecular Evolution and Microbiology		50	4	
SPECIAL PAPER BASED PRACTICAL					
ZOO 396A	FISHERY PRACTICAL –I & Field trip		50	4	
ZOO 396B	ECOLOGY PRACTICAL –I & Field trip				
ZOO 396C	GENETICS & MOL. BIOLOGY PRACTICAL –I & Institute /Lab visit				
ZOO 396D	PARASITOLOGY PRACTICAL –I & Institute /Lab visit				
<b>TOTAL</b>			<b>300</b>	<b>24</b>	
IV	ZOO 401	Environmental pollution & management and Biostatistics		50	4
		ZOO 401.1	Environmental pollution & management		
		ZOO 401.2	Biostatistics		
	ZOO 402	Developmental Biology and Neuroendocrinology		50	4
		ZOO 402.1	Developmental Biology		
		ZOO 402.2	Neuroendocrinology		
	SPECIAL PAPER (A:Fishery; B:Ecology; C:Genetics & Mol. Biology; D:Prasitology)				
	ZOO 403A	Aquaculture & Fish Technology and Inland & Marine Fisheries			
		ZOO 403A.1	Aquaculture & Fish Technology	25	2
		ZOO 403A.2	Inland & Marine Fisheries	25	2
	ZOO 403B	Systems Ecology & Human Ecology			
		ZOO 403B.1	Systems Ecology	25	2
		ZOO 403B.2	Human Ecology	25	2
	ZOO 403C	Human Disease & Molecular Analysis and Applied Genetics			
ZOO 403C.1		Human Disease & Molecular Analysis	25	2	
ZOO 403C.2		Applied Genetics	25	2	
ZOO 403D	Vector Biology & Vector borne Parasites and Molecular Diagnosis & Clinical Parasitology				
	ZOO 403D.1	Vector Biology & Vector borne Parasites	25	2	
	ZOO 403D.2	Molecular Diagnosis & Clinical Parasitology	25	2	
ZOO 494	Environmental Management, Biostatistics, Developmental Biology & Neuro-endocrinology (Practical)		50	4	

SPECIAL PAPER BASED PRACTICAL			
ZOO 495A	FISHERY PRACTICAL –II	50	4
ZOO 495B	ECOLOGY PRACTICAL –II		
ZOO 495C	GENETICS & MOL. BIOLOGY PRACTICAL –II		
ZOO495D	PARASITOLOGY PRACTICAL –II		
SPECIAL PAPER BASED PROJECT/DISSERTATION			
ZOO 496A	FISHERY SPECIAL Pr.	50	4
ZOO 496B	ECOLOGY SPECIAL Pr.		
ZOO 496C	GENETICS& MOL. BIO SPECIAL Pr.		
ZOO 496D	PARASITOLOGY SPECIAL Pr.		
<b>TOTAL</b>		<b>300</b>	<b>24</b>
<b>GRAND TOTAL</b>		<b>1200</b>	<b>96</b>

### M.Sc. ZOOLOGY SEMESTER MARKS DISTRIBUTION

SEM	Theory	Practical
I	200	100
II	200	100
III	200	100
IV	150	150
<b>Total</b>	<b>750</b>	<b>450</b>

Practical papers	
<b>ZOO-195</b>	<b>Marks (Total=50)</b>
Non- Chordates	10
Chordates	12
Histochemistry	12
Animal Physiology	06
Internal assessment- (Viva & LNB)	10
<b>ZOO-196</b>	<b>Marks (Total=50)</b>
Immunology	10
Methods in Biology	05
Cytogenetics	15
Cell Biology	10
Internal assessment- (Viva & LNB)	10

<b>ZOO-295</b>	<b>Marks (Total=50)</b>
Biosystematics	05
Ecological principles	15
Biophysics	05
Biochemistry	15
Internal assessment- (Viva & LNB)	10
<b>ZOO-296</b>	<b>Marks (Total=50)</b>
Molecular Biology	15
Parasitology	15
<b>Field Study</b>	10
Internal assessment- (Viva & LNB)	10

<b>ZOO-395</b>	<b>Marks (Total=50)</b>
Entomology	10
Ecotoxicology	13
Microbiology	17
Internal assessment- (Viva & LNB)	10
<b>ZOO-396 (Special paper)</b>	<b>Marks (Total=50)</b>
ZOO-396A Fishery	30
ZOO-396B Ecology	30
ZOO-396C Genetics & Mol. Bio.	30
ZOO-396D Parasitology	30
Field trip/Institute /Lab visit	10
Internal assessment- (Viva & LNB)	(10) for each special paper

<b>ZOO-494</b>	<b>Marks (Total=50)</b>
Biodiversity and Environmental stress	05
Biostatistics	15
Developmental biology	12
Neuroendocrinology	8
Internal assessment- (Viva & LNB)	10
<b>ZOO-495( Special paper)</b>	<b>Marks (Total=50)</b>
ZOO-495A Fishery	40
ZOO-495B Ecology	40
ZOO-495C Genetics & Mol. Bio.	40
ZOO-495D Parasitology	40
Internal assessment- (Viva & LNB)	10

<b>ZOO-496(PROJECT/DISSERTATION)</b>	<b>Marks (Total=50)</b>
Special paper	
ZOO-496A Fishery	50
ZOO-496B Ecology	50
ZOO-496C Genetics	50
ZOO-496D Parasitology	50

# 1<sup>st</sup> YEAR

## SEMESTER- I

**Course No: ZOO 101: Non-Chordate & Chordates**

**(50 Marks, 4 credit)**

**UNIT I: ZOO 101.1: Non-chordates**

### **Course Outcome:**

*At the end of the course, the students will be acquainted with the non-chordate biology. Students will be able to identify the invertebrates and classify them. Non-chordates (Metazoa) representing largest groups of animal kingdom are characterised by several unique features and display various phenomena (polymorphism, torsions, matamorphosisetc), based on which an array of theories, hypothesis, scientific principles have been advocated in the gamut of the subject of Zoology. The syllabus has provided more stresses on three different aspects:--1) Evolution based on existing theories and hypothesis explaining non-chordate, especially metazoans origin and evolution; 2) Phylogenetic relationships among metazoans based on superphyletic concepts; and 3) Conservation strategies of metazoans for the ecological and economic benefits.*

### **Syllabus:**

1. Origin & Evolution of Metazoa; Phylogenetic overview of major invertebrate phyla.
2. Comparative account about different larval forms of coelomate non-chordates.
3. Biology of the free living nematods – feeding mechanisms and role of nematodes in ecosystem.
4. Bryozoa – anatomical peculiarities feeding mechanisms and phylogenetic relationship.
5. Rotifera – general organisation, mastax, reproduction and cyclomorphosis.
6. Foraminifera – characteristics, origin, distribution, biology and ecological role of foraminifera.
7. Conservation strategies of invertebrates: invertebrate diversity, importance and threats; alternative approaches to species focused conservation; conservation status evaluation for invertebrate species.

### Reference Books/ Journal Article:

1. Animal Evolution-Interrelationships of the living Phyla: Claus Nielsen (Oxford University Press)
2. Diversity of Life (Invertebrates): Harry D. Rounds (East-West Press Pvt. Ltd.)
3. Assembling the Tree of Life (Edited by Joel Cracraft and Michael J. Donoghue (Oxford University Press)
4. An introduction to the invertebrates: Janet Moore (Cambridge University Press)
5. The History of Life -A very short introduction: Michael J. Benton (Oxford University Press)
6. Invertebrate structure and function: Barrington E J W, Thomas Nelson and Sons Ltd, London
7. Invertebrate Zoology: Ruppert and Barnes
8. Biology of the Invertebrates: J A Pechenik
9. Invertebrate Zoology: Anderson
10. Invertebrate Zoology: Meglitsch and Schram
11. Pat Willmer: Invertebrates Relationships

### UNIT II: ZOO 101.2: Chordates

#### Course Outcome:

*Through the course, the students will be accustomed with the chordate biology. Students will be able to identify the chordates and classify them.*

#### Syllabus:

1. **Origin of Chordates:**  
Hemichordata, Cephalochordata, Urochordata, Origin of craniates, Evolution of primates with special reference to *Homo sapiens sapiens*.
2. **Protochordates :**  
Endostyle and Iodine binding capacity in Protochordates.
3. **Fishes:**  
Taxonomy of Fishes, Inland and Marine fisheries of India, Problems & Prospects.
4. **Respiratory system & Gas bladder:**  
General functional and requirements; ventilation of Internal gills; Agnathoxs, Cartilaginous fishes, Bony fishes, larval gills; arial respiration in long fishes; swim bladder and the origin of lungs, lung and other ducts, evolution.
5. **Excretory System and Osmoregulation:**  
General nature of kidneys; Evolution of kidneys, Kidney structure in relation to Osmoregulation; Basic pattern and the Archinephros, Pronephros, Mesonephros,



Metanephros: External salt excretion, Osmoregulation in freshwater and marine water fishes; Association of Urinary System & General system.

6. **Echolocation:**

General consideration of organs of hearing balance and Echolocation; Morphological adaptation for echolocation. Bat Echolocation.

**Reference Books/ Journal Article:**

1. Wolff, R. G. (1991). Functional chordate anatomy, D. C. Heath Canada, Limited. The University of Michigan.
2. PANDEY, B. N. and V. MATHUR (2018). BIOLOGY OF CHORDATES. PHI Learning Pvt. Ltd., 2018.
3. Satoh, N. (2016). Chordate Origins and Evolution: The Molecular Evolutionary Road to Vertebrates, Elsevier Science.Academic Press, 2016
4. Kardong (2005). Vertebrates, 4/E, McGraw-Hill Education (India) Pvt Limited.

**Course No: ZOO 102: Histochemistry & Animal Physiology**

**(50 Marks, 4 credit)**

**UNIT I: ZOO 102.1: Histochemistry**

**Course Outcome:**

*Students will understand the Scope and importance of histology and histochemistry. General principles for the preparation of tissue for histological studies and histochemical localization of various cellular components would be another significant aspect of the course*

**Syllabus:**

1. Introduction to Microtechnique
2. Fixation
3. Dyes
4. Histological staining
5. Enzyme histochemistr
6. Immunohistochemistry

**Reference Books/ Journal Article:**

1. Bancroft, JD (1975) Histochemical Techniques (Second Edition), Butterworth-Heinemann.
2. Kirenan, JA (1981) Histological and Histochemical Methods, Theory and Practice (Third Edn.)
3. Suvarna, SK, Layton, C, Bancroft, JD (1977) Bancroft's Theory and Practice of Histological Techniques. Churchill Livingstone.

**UNIT II: ZOO 102.2: Animal Physiology**

## **Course outcome:**

*The course will provide detailed knowledge on the various physiological organ-systems and their importance to the integrative functions of the human body. To have an enhanced knowledge and appreciation of mammalian physiology; to understand the functions of important physiological systems including the cardio-respiratory, renal, reproductive and metabolic systems; to understand how these separate systems interact to yield integrated physiological responses to challenges such as exercise, fasting and ascent to high altitude, and how they can sometimes fail; to be able to perform, analyze and report on experiments and observations in physiology; to be able to recognize and identify principal tissue structures.*

## **Syllabus:**

### 1. Blood, Circulation and Respiration:

- Haemopoiesis, haemoglobin, blood groups, haemodynamics.
- Regulation of blood volume and blood pressure, haemostasis.
- Respiratory response to extreme conditions like hypoxia & diving.
- Body oxygen stores – blood, muscle and pulmonary.
- Oxyhaemoglobin and Myoglobin; Oxygen dissociation curve.
- Cardiovascular System:
  - Cardiac cycle,
  - Electrical and mechanical properties of myogenic and neurogenic hearts;
  - Heart as a pump; regulation of heart pumping;
  - Neural and chemical regulation of excitation & conduction in heart;
  - Frank-Starling mechanism;
  - Principle of ECG.

### 2. Stress physiology:

- Homeostasis, Feedback control systems
- Oxidative stress. Cellular response. Free radicals and anti-oxidants.

### 3. Thermoregulation:

- Body temperature and determinants of body heat – production and loss.
- Physiological events for thermoregulation; counter-current system.
- Thermal biology of ectotherms, heterotherms and endotherms.

## **Reference Books/ Journal Article:**

1. Textbook of Medical Physiology - Arthur C. Guyton & John Edward Hall. 13<sup>th</sup> Ed.
2. Ganong's Review of Medical Physiology- Kim E. Barrett, Susan M. Barman, Scott Boitano, Heddwen Brooks. 25th Ed.
3. Biochemistry - Debajyoti Das, 1978.
4. A Textbook of Practical Physiology- C. L. Ghai. 2012.

## **Course No: ZOO 103: Immunology and Methods in Biology**

**(50 Marks, 4 credit)**

### **UNIT I: ZOO 103.1: Immunology**

#### **Course outcome:**

*This course will describe the immune systems of vertebrates that enable them to recognize and respond specifically to foreign substances. The students will be able to comprehend the roles of lymphoid organ, cells of immune system, antigens, antibodies, MHC, antigen presentation and immunity to infectious diseases.*

#### **Syllabus:**

1. a) Cells and organs involved in Immune System, Types of Immunity
2. a) Antigenicity and Immunogenicity  
b) Concept of Epitope, Paratope, Agreptope, Hapten and Adjuvants
3. a) Origin and maturation of T and B lymphocyte, Humoral and cell mediated Immune Response  
b) T-cell subpopulation
4. a) Antigen processing and presentation  
b) Major Histocompatibility Complex (MHC) Mechanism of immune response and generation of immunological diversity
5. Complement system: classical, alternative and lectine pathway, MAC formation and related disorders
5. a) Structure and function of Immunoglobulin (Ig) and its Isotypes.  
b) Enzymatic activity on Ig molecule.
6. Applied Immunology:-  
a) ELISA, RIA, Southern blotting hybridization, Immunohistochemistry

#### **Reference Books/ Journal Article:**

1. Abbas, A. K., Lichtman, A. H. and Pillai, S. (2006). *Cellular and molecular Immunology*. 6<sup>th</sup> ed. Saunders.
2. Abbas, A. K. and Lichtman, A. H. (2006). *Basic Immunology*. 2<sup>nd</sup> ed. Elsevier.
3. Coico R, Sunshine, G., Benjamini, E. (2003). *Immunology: A short Course*. 5<sup>th</sup> ed. Wiley-Liss: New Jersey.

4. English, L. S. (1994). *Technological Applications of Immunochemicals (BIOTOL)*. Butterworth- Heinemann, Oxford Freeman and Co.
5. Goldsby, R. A., Kindt, T. J., Kuby, J. and Osborne, B. A. (2013). *Immunology*. 7<sup>th</sup> ed. W. H. Freeman and Co.
6. Khan F. H. (2009). *The Elements of Immunology*. Prentice Hall India.
7. Kindt, T., Goldsby, R. Osborne, B. (2007). *Kuby's Immunology*. 6<sup>th</sup> ed. W.H. Freeman and Co.
8. Male, D., Brostaff, J., Roth, D. and Roitt, I. (2006). *Immunology*. 7<sup>th</sup> ed. Mosby.
9. Rao, C. V. (2002). *Immunology*. Narosa Publishing House, New Delhi.
10. Roitt, I. M. and Delves, P. J. (2001). *Roitt's Essential Immunology*. 10<sup>th</sup> ed. Blackwell Science Ltd.

## UNIT II: ZOO 103.2: Methods in Biology

### Course Outcome:

*On completion of this course students will be able to: address a research problem in biotechnology, Provide examples of current applications of biotechnology and advances in the different areas like medical, microbial, environmental and bioremediation. After getting theoretical knowledge students would be acquainted with some modern instruments and methods which are indispensable to pursue advance research in the field of biotechnology.*

### Syllabus:

#### 1. Molecular Biotechnology

- a) Recombinant DNA technology
- b) Restriction Endonuclease
- c) Production of recombinant DNA molecule
- d) Cloning Vector
- e) Amplification by PCR
- f) DNA finger printing and its application

#### 2. Environmental Biotechnology

- a) Bioremediation
- b) *In situ* bioremediation
- c) *Ex situ* bioremediation
- d) Bioremediation of Xenobiotic components and hydrocarbons
- e) Phytoremediation
- f) Cryopreservation; Integration of different rural biotechnological tools

#### 3. Techniques and Bioinstrumentation

- a) Principles and application of gel-filtration, ion-exchange and Affinity Chromatography, Thin layer and Gas Chromatography- MS.
- b) Basic Principles of Electrophoresis, Agarose Gel Electrophoresis, SDS-PAGE, Cell fractionation, Ultracentrifugation, , Western Blotting Hybridization.
- c) Flow Cytometry, 2D Gel Electrophoresis, FISH, FTIR.

**Reference Books/ Journal Article:**

1. Principle and techniques of biochemistry and Molecular Biology by K.Wilson and J. Walker 7<sup>th</sup>Edn, Cambridge low price edn.
2. Physical biochemistry- Principles and Applications by David Sheehan, 2<sup>nd</sup> edn.

**Course No: ZOO 104: Cell Biology & Cytogenetics**

**(50 Marks, 4 credit)**

**UNIT I: ZOO 104.1: Cell Biology**

**Course outcome:**

*Students will be well informed with the membrane structure and composition, transport and trafficking of protein, the cytoskeleton, cell movement and extra cellular matrix. The mechanism of cell division and their regulation through different check points will be thoroughly understood. Cell cycle, apoptosis, signal transduction and cancer biology are the important part of the course.*

**Syllabus:**

**1) Biomembrane Structure**

The Lipid Bilayer: Composition and Structural Organization, Membrane Proteins: Structure and Basic Functions, Phospholipids, Sphingolipids, and Cholesterol: Synthesis and Intracellular Movement

**2) Transmembrane Transport of Ions and Small Molecules**

Overview of Transmembrane Transport, ATP-Powered Pumps and the Intracellular Ionic Environment, Overview of Transcellular Transport

**3) Moving Proteins into Membranes and Organelles**

Targeting Proteins to and across the ER Membrane, Insertion of Membrane Proteins into the ER, Targeting of Proteins to Mitochondria and Chloroplasts, Transport Into and Out of the Nucleus

#### 4) **Signal Transduction**

Basic idea of Cell signaling, G Protein–Coupled Receptors: Structure and Mechanism, G Protein–Coupled Receptors and Regulation of Ion Channels, Signaling through second messengers, Receptor tyrosine kinase signaling, MAP Kinase

#### 5) **Cytoskeleton & Cellular Motility**

Microtubule Dynamics and regulation, Microtubular motor proteins: Kinesins&Dyneins and Cellular motility

#### 6) **Cell cycle & its regulation:** A cycle of cyclin dependent kinase activities regulates cell proliferation, Regulation of CDK-cyclin complexes, Protein kinases in cell cycle

#### 7) **Interactions between Cells and Their Environment:**

Overview of major cell-cell and cell-matrix adhesive interactions, Cell-Cell and Cell–Extracellular Junctions and Their Adhesion Molecules, The Extracellular Matrix: The Basal Lamina and Connective tissue

#### **Reference Books/ Journal Article:**

1. Cell And Molecular Biology by Gerald Karp
2. Lewin's Cells by Gorge Plopper, David Sharp
3. Molecular Cell Biology by Harvey Lodish
4. Molecular Biology The Cell by Bruce Alberts
5. The Cell by Geoffrey M. Cooper
6. Molecular Biology by Robert F. Weaver

#### **UNIT II: ZOO 104.2: Cytogenetics**

#### **Course outcome:**

*The course will provide an understanding of genetic analysis at the gene, genome and population levels. Understanding the Drosophila genetics. Evaluation of the various aspects of structural, functional and comparative genomics. Designing and development of experiments using Drosophila and their evaluation through genetic analysis and interpretations*

#### **Syllabus:**

##### 1. **Genetic Fine structure :**

The CIS-TRANS or complementation test for functional allelism, Fine structure of the phage T4 rII locus, Complementation mapping and deletion mapping.

2. **Recombination in Bacteria :**

F factor, episomes, Hfr, integration of F factor, Interrupted mating Experiment, conjugation mapping, transformation and transduction

3. **Tumor Inducing Viruses – Viral Oncogenes**

Life Cycle of Rous Sarcoma Virus, RSV genome organization, mechanism of integration, formation of transducing retroviruses, protein products of protooncogene, Oncoproteins regulate gene expression and signal transduction Cancer induction by Retroviruses, tumor suppressor gene and their function.

4. **Genetic structure of Populations -**

Genotypic frequencies, Allelic Frequencies, the Hardy Weinberg Law, calculation of genotypic and allelic frequencies where multiple alleles are present, derivation the Hardy-Weinberg Law

**Reference Books/ Journal Article:**

1. Introduction to Genetic Analysis by J.F.Griffiths
2. Genes viii by Benjamin Lewin
3. Genetic: Analysis and Principles by Robert J. Brooker
4. An Introduction to the Genetic Analysis by David T. Suzuki
5. Genetics:A Conceptual Approach by Benjamin A.Pierce
6. iGenetics: A Molecular Approach by Peter J. Russell
7. Principle of Genetics by Peter Snustad

**Course No: ZOO 195: Non- Chordates, Chordates, Histochemistry & Animal Physiology (Practical)**

**(50 Marks, 4 credit)**

**Syllabus:**

- 1) Non- Chordates:
  - i) Identification of common Invertebrate and Vertebrate taxa
  - ii) Minor Dissection:
    - (1) Grasshopper - Reproductive system/ Nervous system
    - (2) Cockroach – Stomatogastric Nervous system
    - (3) Achatina – Reproductive system & Nervous system
- 2) Chordates:
  - i) Major Dissection
  - ii) Bony fish - Vth, VIIth cranial nerves

- 3) Histochemistry
  - a. Microtechniques and tissue identification from chordates
  - b. Enzyme histochemistry
  - c. Staining for nucleic acid/ connective tissue
  - d. Demonstration of research article.
  - e. General idea about modern tools of histological techniques.
  
- 4) Animal Physiology
  - i) Estimation of pH and its impact on plankton.
  - ii) Observation of gut movement in frog/rat/fish under hypoxia using Dale's apparatus

**Course No: ZOO 196: Immunology, Methods in Biology, Cell Biology & Cytogenetics (Practical)**

**(50 Marks, 4 credit)**

**Syllabus:**

**1. Immunology:**

- a. Study of macrophage.
- b. Study of phagocytosis.
- c. Determination of human blood group

**2. Methods in Biology**

- a. Characterization of macromolecule through Gel electrophoresis

**3. Cell Biology**

- a. Identification of different stages of cell division and cell organelle.
- b. Mitochondrial Staining

**4. Cytogenetics:**

- a. Life cycle of *Drosophila*.
- b. Analysis and interpretation of genetic crosses with special reference to *Drosophila*
- c. Study of polytene chromosome of *Drosophila*.



# 1<sup>st</sup> YEAR

## SEMESTER- II

**Course No: ZOO 201: Biosystematics & Ecological principles**

**(50 Marks, 4 credit)**

**UNIT I: ZOO 201.1: Biosystematics**

### **Course outcome:**

*This course will describe the importance of taxonomy in biology, historical resume of systematics and stages of taxonomy. The course would be dealt with biological classification, basic principles and rules for the classification of organisms. The course would enlighten the students with modern trends in biosystematics-concepts of different conventional and newer aspects.*

### **Syllabus:**

1. **Microtaxonomy:** Phenon, Taxon, Category, type; stages of taxonomy; Aims and tasks of Taxonomists; Importance of taxonomy in Biology.
2. **Macrotaxonomy:** Theory and practice of Biological classification; Basic principles, Rules for the classification of organisms, Identification criteria, Taxonomic characters, Classification and phylogeny, Is classification a Theory? The functions of a classification.
3. **Concept of Species:** Typological species concept, Nominalistic species concept, Biological species concept, Evolutionary species concept; other kinds of species; Polytypic species, Subspecies, Intraspecies and Superspecies.
4. **Newer Systematics:** Morphological approach, Immature stages and Embryological approach, Ecological approach, Behavioural approach, Ecological approach, Behavioural approach, Cytological approach, Biochemical approach, Numerical systematics, Differential systematics.
5. **Molecular Systematics:** Immunological aspect, chromatographic aspect, Electrophoresis, Infrared spectrophotometry, Histochemical studies, genetic complement, DNA hybridization, Karyological studies.
6. **Macromolecular & Micromolecular Systematics:** based on DNA, RNA, Protein, amino acids, fatty acids and phenols.
7. **Role of Systematics in applied Biology:** Agriculture & Forestry, Biological control, wild life management, National defence, Environmental problems, soil fertility, Mineral prospecting, Quarantine measure, Commercial application.
8. **Systematics and Public Health Management**

**Reference Books/ Journal Article:**

1. Mertens, T. R. and J. L. Lines (1978). Principles of biosystematics, Educational Methods.
2. Daniel, M. (2009). Taxonomy: Evolution at Work, Alpha Science International. Publisher Alpha Science International, 2009
3. Mayr, E. and P. D. Ashlock (1991). Principles of systematic zoology, McGraw-Hill. Publisher McGraw-Hill, 1991
4. Hickman, C. P., S. L. Keen, et al. (2016). Integrated Principles of Zoology, McGraw-Hill Education. 17th Eds.

**UNIT II: ZOO 201.2: Ecological principles****Course outcome:**

*The subject ecology dealing with the relationship among different life forms in respect of their environment is a blend of concepts and contents (information). In order to understand the applicability of this subject, the students must have an acquaintance with the different definitions, terminologies, scientific principles, hypothesis, theories etc. with proper examples. The contents of this syllabus have been identified with an eye to make the students understand the basics of ecology putting more emphasis on the system ecology , habitat ecology, population and community ecology and evolutionary ecology so that ecology students can not only understand the ongoing ecological processes but also can contribute to the proper eco-management.*

**Syllabus:****1. Basics of Ecology**

Biosphere and Ecosphere; Types of food web : Connectedness, energy and functional webs; Features of food web – nodes, links, linkage density, connectance, chain length; cybernetic nature of ecosystem; stability through feedback control and through redundancy of components; resistance and resilience stability, Gaia hypothesis.

**2. Population Ecology**

Survivorship; Life table, fertility schedule. Reproductive strategies; semelparity, iteroparity, r & k strategies, population interactions- direct and indirect, positive and negative. Lotka-volterra model of competition and predator-prey interaction. Causes of extinction and endangerment of populations. Anthropogenic impact on species extinction, habitat destruction and fragmentation, introduction of exotic species.

**3. Community and Ecosystem**

Structure of biotic community. Community patterns: diversity and stability. Community boundary: Ecotone and edge types, Edge effect and edge species, Edge/Area ratio in relation to size, shape and fragmentation of habitat. Organismic and individualistic concepts of community. Leibig's Law of tolerance.

#### **4. Habitat Ecology**

Habitat and niche: spatial, trophic and multi-dimensional niche concepts, fundamental and realized niche, niche breadth and niche overlap. Competitive exclusion: experimental and natural evidence. Keystone species. Foundation species. Species abundance hypothesis. Ecological guilds and ecological equivalents.

#### **5. Evolutionary Ecology**

Definition; different approaches. Bet-Hedging strategies. Hamilton's role and limitations of inclusive fitness model.

#### **Reference books:**

- Fundamentals of Ecology- Eugene P. Odum, 2005.
- Biological Science – Scott Freeman. 2018.
- Ecology- Robert E. Ricklefs, Gary Leon Miller. 2000.

### **Course No: ZOO 202: Biophysics & Biochemistry**

**(50 Marks, 4 credit)**

#### **UNIT I: ZOO 202.1: Biophysics**

#### **Course outcome:**

*At the end of the course the student will be able to understand fundamental concepts in biophysics that underlie biological processes. Thorough knowledge of biophysics of membrane and laws of thermodynamics. The course would be dealt with principle of thermodynamics, electromagnetic and ionizing radiation and principles of biophysical chemistry.*

#### **Syllabus:**

1. Biophysical principles.
2. Thermodynamics
3. Coloidal system
4. Microscopy in biology and medicine

5. Biophysics of membrane
6. Dynamics of circulation

### Reference books:

1. Cotterill, R. (2005) Biophysics: An Introduction. Jhon Wiley & Sons.
2. Bialek, W. Biophysics: Searching for Principles.
3. Cleri, F. (2016) The Physics of Living System. Springer.

### UNIT II: ZOO 202.2: Biochemistry

#### Course outcome:

*The course will provide an understanding of fundamental biochemical principles such as biomolecules, metabolic pathway and regulation of biological process. On completion of the course, students are able to: Understand about the agencies responsible for Production of various products using biochemistry. Understand the term pH, Buffer. Understand the structure and function of carbohydrate, amino acids, proteins, and lipids. The students will understand the fundamental energetic of biochemical processes and chemical logic of metabolic pathways. The course also deals with the integration of metabolic process in cellular systems and organization of cellular pathways.*

#### Syllabus:

1. **Stablizing** interactions (Van der Waals, electrostatic, hydrogen bonding, hydrophobic interaction)
2. **Protein Conformation:** Primary, secondary, tertiary and quaternary structures; Ramachandran plot; domains; motif and folds.
3. **Enzymes:** Enzyme kinetics, Michaelis-Menton equation, hyperbolic and Lineweaver-Burke plot; co-enzymes and Cofactor; competitive and non-competitive inhibitor and their effects on enzyme kinetics; Active site of an enzyme; Enzyme regulation, allosteric modification, its kinetics, covalently modulated enzymes.
4. **Biological Oxidation:** Redox potential, mitochondrial electron carriers, the respiratory chain (electron transport chain); Mitchell's chemiosmotic theory of oxidative phosphorylation; FoF, ATPase
5. **Lipid Metabolism:** denovo synthesis of fatty acids, microsomal fatty acid elongase&desaturase systems; oxidation of saturated fatty acids and unsaturated fatty acids.

6. **Protein metabolism:** deamination, transamination, ammonotelism, ureotelism, uricotelism, formation of urea, formation of specialized products from amino acids: catecholamine, serotonin, melatonin, glutathione, T<sub>3</sub>, T<sub>4</sub>.
7. **Carbohydrate metabolism:** anabolic role of TCA cycle, integration of carbohydrate, fat and protein metabolism. Regulation of Glucolysis TCA cycle, Gluconeogenesis, Pentose phosphate pathway, Glycogenesis, glycogenolysis with special reference to rate limiting steps.

#### Reference books / Journal articles:

1. Biochemistry by Jeremy M. Berg, John L. Tymoczko, Lubert Stryer
2. Biochemistry by D Voet and J G. Voet
3. Biochemistry by Mary K. Campbell, Shawn O. Fa
4. Lehninger Principles of Biochemistry by David L. Nelson and Michael M. Cox
5. Biochemistry by David E. Metzler

### Course No: ZOO 203: Molecular Biology & Parasitology

(50 Marks, 4 credit)

#### UNIT I: ZOO 203.1: Molecular Biology

#### Course outcome:

*The course will be able to explain the fundamentals of genetics and the Mendelian laws, the concept of alleles, concept of linkage and crossing over of genes. The course will open an avenue to be familiar with a variety of types of genetic data (genotyping, expression, and sequence data), chromosomal mapping, genetic composition of biological population and evolutionary factors that explain the variation. An in-depth knowledge of chemical and molecular processes that occur in between cell including the central dogma will be assured at the end of this course.*

#### Syllabus:

##### 1. DNA Replication:

The chemistry of DNA synthesis, the mechanism of DNA polymerase, the replication fork, the specialization of DNA polymerase, finishing replication

##### 2. The Transcription Process –

Role of RNA polymerase in prokaryotes, initiation of transcription at Promoters, elongation and termination of an RNA in prokaryotes, Initiation, elongation, DNA methylation and control of transcription in eukaryotes

##### 3. Protein Synthesis

Charging tRNA, initiation of translation; role of initiation factors, Elongation: binding of Aminoacyl-tRNA, peptide bond formation and translocation. Termination of translocation.

#### **4. Regulation of Gene Expression in Prokaryotes:**

The Operon Model; lac, an inducible Operon, Positive Control of the lac Operon by CAP and Cyclic AMP. Repressible operon, Gene organization of the Tryptophan biosynthesis gene, Regulation of the trp operon.

#### **Reference books / Journal articles:**

1. Molecular Biology of the Gene by James D. Watson
2. Genes viii by Benjamin Lewin
3. Genetics: A Conceptual Approach by Benjamin A. Pierce
4. iGenetics: A Molecular Approach by Peter J. Russell
5. Principle of Genetics by Peter Snustad
6. Concept of Genetics by William S. Klug, Michael R. Cummings
7. Introduction to genetics A Molecular Approach by Terry Brown
8. Molecular Biology by Robert F. Weaver

## **UNIT II: ZOO 203.2: Parasitology**

### **Course outcome:**

*The course will provide an understanding of the diversity and biology of parasites, besides the epidemiological aspects of different parasitic diseases would be explored and students will be able to gain knowledge regarding the mode of transmission of parasitic diseases and preventive measures.*

1. a) Types of Parasites and hosts.  
b) Basic concept of Parasitism, symbiosis, phoresis, commensalisms and mutualism.
2. Molecular, cellular and physiological basis of host-parasite interactions.
3. Life cycle and immunology of *Plasmodium falciparum*, African Trypanosomiasis.
4. Epidemiology and transmission of parasitic diseases. Malaria, Kalazar, Filaria.
5. a) Zoonosis and Zoonotic diseases with special reference to Balantidiasis, Giardiasis, Filariasis and Paragonimiasis.  
b) Life cycle and biology of *Leishmania*, *Schistosoma*.
6. Structure and composition of helminthes cuticle.
7. Vector biology with special reference to mosquito/Sand fly/ticks.

#### **Reference books:**

1. Bogitsh, B. J. and Cheng, T. C. (2000). *Human Parasitology*. 2nd Ed. Academic Press, New York.
2. Chandler, A. C. and Read. C. P. (1961). *Introduction to Parasitology*, 10th ed. John Wiley and Sons Inc.
3. Chatterjee, K. D. (1981). *Parasitology (Protozoology and Helminthology)*. 13<sup>th</sup> ed. CBS.
4. Cheng, T. C. (1986). *General Parasitology*. 2nd ed. Academic Press, Inc. Orlando. U.S.A.
5. Cox, F. E. G. (1993). *Modern Parasitology*. 2nd ed. Blackwell Scientific Publications. Lea and Febiger, Philadelphia.
6. Hati, A. K. (2001). *Medical Parasitology*. Allied Book Agency, Kolkata.
7. Noble, E. R. and Noble G. A. (1989). *Parasitology. The Biology of animal Parasites*. 6th ed. Lea and Febiger, Philadelphia.
8. Roberts, L. S., Janovy, J. and Nadler S. (2013) *Gerald D. Schmidt & Lary S. Roberts' Foundation of Parasitology*. 9<sup>th</sup> ed. McGraw-Hill International.
9. Schmidt, G. D. and Roberts, L. S. (2001). *Foundation of Parasitology*. 3rd ed. McGraw Hill Publishers.
10. Schmidt, G. D. (1989). *Essentials of Parasitology*. Wm. C. Brown Publishers (Indian print; 1990, Universal Book Stall).
11. Smyth, J. D. (1994). *Animal Parasitology*. 3rd ed. Cambridge University Press.

**Course No: C-ZOO 204: Wildlife & Eco-management and Aqua informatics  
(CBCS)**

**(50 Marks, 4 credit)**

**UNIT I: C-ZOO 204.1: Wildlife & Eco-Management**

**Course outcome**

*The syllabus of this subject includes some pertinent aspects on biodiversity dealing with identifying the threats and conservation of biodiversity alongside highlighting some basic aspects of wildlife and its conservation with special reference to the diversity, distribution and conservation of wildlife of South West Bengal, India. This course is expected to generate interest to the students across the disciplines so that they can become a part of wildlife and biodiversity conservation.*

**Syllabus:**

- a) Wildlife diversity and distribution in India – Mammals, Avifauna and Reptiles; IUCN threatened categories.
- b) Threatened wildlife in India with special reference to Eastern India.
- c) Root causes of depletion of wildlife wealth.
- d) Wildlife conservation strategies-Protected areas-National Parks, Sanctuaries, Biosphere Reserve; Cores and Buffer; Nodes and Corridor.

- e) Management of wildlife-Taxonomic Status, Distribution, Habitat Utilization Patterns, Threats and conservation of Mask Deer; Vultures; Olive Ridley turtle.
- f) Tools and Techniques-  
PRA methods ; Molecular Techniques; Tele satellite images; Radio coloring ; Peoples Participation; Ground truth Assessment-Pugmarks, Call counts, Capture-Recapture.  
Wild life trades, Crimes, Laws & Ethics.
- g) Environment –different relevant terminology.
- h) Environmental Management-Basic steps Sustainability, Ecomonitoring Impact Assessment.
- i) Pollution-Types: Mode of action and Environmental Consequences; Global Scenario.
- j) Conservation Biology –different concepts and approaches.

**Reference books / Journal articles:**

1. An Introduction to Disturbance Ecology-A road map for Wildlife Management and Conservation( Springer)
2. Wildlife in India: B Saharaia
3. A manual for wildlife Radio Tagging by Robert E Kenward
4. Conservation of Exploitedspecies(Edted by John D. Renolds, GeorginaM. Mace, Kent H. Redford and John G. Robinson) (Cambridge University Press)
5. Ecological Census techniques: Sutherland
6. Wild life Biology: R F Dasnan
7. A manual of wild life conservation: R D Teague
8. Animal Behavior: A synthesis of ethology and comparative psychology: RA Hinde
9. Animal behavior: Dugartkin
10. Animal behavior: Alcock

**UNIT II: C-ZOO 204.2: Aqua informatics**

**Course outcome:**

*Aquaculture is now viewed as a strong option to increase fish production as it plays a vital role in providing food security in India. Aquaculture informatics can be described as the scientific application of Information technology in biological concepts that enhance the productivity and economic viability of aquaculture sectors. Advances in electronic communication, combined with specific cooperative efforts, should be used to increase the information exchange in the region. It is nation's part to educate the fish farmers through community based organization and to provide an information system, in low cost, to fulfill their requirements.*



## **Syllabus:**

1. Spatial database development through different survey information analysis, Decision base support system formation.
2. Climate change and policy research design on Aquatic resources from beginning to end. Role of Information Communication Technology (ICT) in Aquaculture sector.
3. Recent approaches in Aquatic floral -faunal conservation/ assessment through Web based system.
4. Technology Innovation and Integrated Information Management System (software Program) in Aquaculture.

### **Reference books / Journal articles:**

1. Roy, A. K. and N. Sarangi (2008). Applied Bioinformatics, Statistics & Economics in Fisheries Research, New India Pub. Agency.
2. Burnell, G. and G. L. Allan (2009). New Technologies in Aquaculture: Improving Production Efficiency, Quality and Environmental Management, CRC Press.
3. Food and A. O. o. t. U. Nations (1964). FAO Fisheries Technical Paper, Food and Agriculture Organization of the United Nations (ISBN 9251055548, 9789251055540).
4. Zazueta, F. S. (2006). Computers in Agriculture and Natural Resources: Proceedings of the 4th World Congress: Orlando, Florida, USA, July 24-26, 2006, American Society of Agricultural and Biological Engineers.
5. Bank, T. W. (2012). Agricultural Innovation Systems: An Investment Sourcebook, World Bank Publication. (ISBN 0821389440, 9780821389447). Length 680 pages.

## **Course No: ZOO 295: Ecological principles, Biochemistry & Field Study)**

**(50 Marks, 4 credit)**

### **Syllabus:**

#### **1. Biosystematics**

- i) Preparation of taxonomic key

#### **2. Ecological principles**

- a. Estimation of primary productivity in aquatic ecosystems
- b. Estimation of transparency of water
- c. Measurement of intensity of light – using Lux meter.
- d. Determination of the minimum size and number of quadrat – Species area curve method.
- e. Study of density, diversity, frequency and abundance of plant community.

#### **3. Biophysics**

- a. Membrane biology dynamics
- b. USIC/ SIF visit for Lab Demonstration
- c. Demonstration of Scientific Techniques using local Species as an experimental tool

#### **4. Biochemistry**

- a) Quantitative estimation of protein- Lowry method or by FolinCiocalteu reagent.
  - b) Estimation of Glucose by Dinitrosalicylic (DNS) acid reagent.
  - c) Estimation of Fructose by Resorcinol reagent.
  - d) Estimation of DNA by Diphenylamine reagent.
  - e) Detection of reducing sugars by Benedict's, Barfoed's& Fehling's reagents.
  - f) Detection of amino acids by Ninhydrin reaction.
  - g) Determination of Km &Vmax of enzymes Amylase and/or Alkaline phosphatase.
- Preparation of Progress Curve of the above mentioned enzymes.

**Course No: ZOO 296: Biosystematics, Molecular Biology, Parasitology & Biophysics**  
**(50 Marks, 4 credit)**

**Syllabus:**

1. Parasitology
  - a) Smear preparation and staining of rectal content of Bufo sp./Cockroach
  - b) Preparation and staining of blood parasite from pigeon blood.
  - c) Identification:  
*Plasmodium sp., Leishmania sp., Ascaris sp., Fasciola sp., Paramphistomum sp., Anopheles sp., Culex sp., Aedes sp. Columbicola sp., Pediculus sp., Cimex sp.*
2. Molecular Biology
  - a) Isolation & purification of DNA from tissue.
  - b) Principle & method of Agarose Gel Electrophoresis
3. Field report & viva

# 2<sup>nd</sup> YEAR

## SEMESTER- III

**Course No: ZOO 301: Basic & Applied Entomology and Ecotoxicology**

(50 Marks, 4 credit)

**UNIT I: ZOO 301.1: Basic & Applied Entomology**

### **Course outcome:**

*The subject entomology dealing with the study of different aspects of insects, the largest animal taxa of the world has been included in the general syllabus of Zoology in third semester mainly to give the students an understanding on three different aspects of entomology – morphology and classifications, insects' physiology and some unique phenomena in the insects life such bioluminescence, pheromones, secondary adaptation aquatic insects, insects-plants interactions , integrated management of insects pests etc. so that the students can develop not only the interest on this interesting and important faunal components but can undertake measures for the conservation of beneficial insects and control of the harmful ones.*

### **Syllabus:**

1. **The importance, diversity and conservation of insects** – Insect biodiversity, uniqueness and adaptability, insect conservation. Insect for food and silk – prospects and problems of sericulture in drought prone lateritic tracts of South West Bengal, India.
2. **General characters and classification of Insects up to order** - Insect's head, capsule, antennae, legs, wings, digestive system with special emphasis to midgut, filter chamber and peritrophic membrane; integument, Insects' neuro-endocrine system – components, chemical structure of hormones and functions; molting and metamorphosis, insects' egg-type, hatching, growth, development, diapause and aestivation.
3. **Biology, nature of damage and control of Insects' pests** - Jute, cashew, betel vine and stored grains; Integrated approach to pest management.
4. **Aquatic insects** – Diversity of freshwater and marine insects, Adaptation – water balance; Importance for environmental monitoring.
5. **Insect behaviour** - Pheromones – Structure of pheromone glands; types and functions; biochemical synthesis of pheromones. Bioluminescence – Light producing organs, Mechanism of light production, Control and significance of light production.

6. **Insects and Plants** – Insect plant interaction and co-evolutionary interactions between plants and animals; Plant chemicals and their effect on insects; Pollination by insects; Organic compounds and their biosynthesis pathways in insects

**Reference books / Journal articles:**

1. Principal of Insect morphology R E Snodgrass
2. Imms general Text Book of Entomology O W Richards & R G Davies
3. The Insects: Structure & Function R F Chapman
4. General and Applied Entomology K KNayar, Ananathakrishnan& David
5. The science of Entomology Romser&Stoffolans
6. Insect Pest Management :D Dent
7. Entomology & Pest management L P Pedigo
8. Insect Pests in Tropical Forestry: Martin R.Speight( CABI Publishing)
9. Entomology: C Gillot
10. Insect evolutionary ecology: M D E Fellowery, G J Holloway
11. Insect Physiological Ecology-Mechanisms and Patterns: Steven L. Chown and Sue W. Nicolson( Oxford University Press)
12. Pheromones and Animal Behaviour: TristramD.Wyatt( Oxford University Press)

**UNIT II: ZOO 301.2: Ecotoxicology**

**Course outcome:**

*The purpose of toxicity testing is to generate information about a substance's toxic properties so that the health and environmental risks it poses can be adequately evaluated. In aquatic sector toxicological study may ensure the health of the local aquatic organism and related to human health.*

**Syllabus:**

**1. Xenobiotics**

General idea of Xenobiotics and their Physical & Chemical Properties; Corrosive, Metabolic, Neurotoxic, Mutagenic & Carcinogenic toxins; Characteristics of toxin, Route of Entry, Mechanism of Action.

**2. Toxicity test & bioassay**

LC<sub>50</sub>, LD<sub>50</sub>, Dose response curve; Biotransformation, Bioaccumulation & Biomagnification of Xenobiotics in food chain; Hazardous heavy metals and their toxicity and probable antidotes; Elementary idea on Chelation therapy.

**3. Aquatic Toxicology**

A short history of Aquatic toxicology, The aquatic environment, Factors affecting the Environmental Contraction of Chemicals, Toxicological Concept and Principles, Factors influence Toxicity, Toxic agents and their effects, concentration – Response Relationships, toxicity testing, Biomonitoring Toxicity data and Environmental regulation.

#### 4. Immunotoxicology

Immunology – Defensive responses, Immunological methodology; Immunotoxicology – Effects of classes of Toxicants.

#### 5. Environmental Genotoxicology

Basic mechanism of DNA damage, Analytical techniques, In situ Environmental Genotoxicity studies with Aquatic species, potential value of Environmental genotoxicity.

#### Reference books / Journal articles:

1. Newman, M. C. and W. H. Clements (2007). Ecotoxicology: A Comprehensive Treatment, CRC Press. CRC Press, 2007.
2. Jorgensen, E. (2010). Ecotoxicology, Elsevier Science. Academic Press, 2010.
3. Walker, C. H., R. M. Sibly, et al. (2016). Principles of Ecotoxicology, Fourth Edition, CRC Press.
4. Hoffman, D. J., B. A. Rattner, et al. (2002). Handbook of Ecotoxicology, Second Edition, CRC Press.
5. Forbes, T. L. (1993). Ecotoxicology in Theory and Practice, Springer Netherlands. Springer Science & Business Media, 1993.

### Course No: ZOO 302: Molecular Evolution and Microbiology

(50 Marks, 4 credit)

#### UNIT I: ZOO 302.1: Molecular Evolution

#### Course outcome:

*The curriculum in evolution includes modern aspects of evolution and molecular phylogenetic tree. DNA and amino acid sequences study by genomics and proteomics data analysis and find out the similarities and dissimilarities between different species of same group or different group. By nearest neighbor analysis and by principle of parsimony a gene tree could be made. PG students are benefitted by study of genome analysis and by finding the sequences they may be able to find out the molecular systematic position.*

#### Syllabus:

##### 1. Neo-Darwinism

- a. Hardy-Weinberg law of genetic equilibrium
- b. A detailed account of destabilizing forces: (i) Natural selection (ii) Mutation  
(iii) Genetic drift (iv) Migration

##### 2. Molecular phylogenies

- a. Construction of phylogenetic trees

- b. Phylogenetic inference-Distance methods, parsimony methods, maximum likelihood method
- c. Immunological techniques
- d. Amino acid sequences and phylogeny
- e. Nucleic acid phylogeny, DNA-DNA hybridizations, restriction enzyme sites, nucleotidesequence comparisons and homologies

**Reference Books/ Journal Article:**

1. Introduction to Genetic Analysis by J.F.Griffiths
2. Genetic: Analysis and Principles by Robert J. Brooker
3. An Introduction to the Genetic Analysis by David T. Suzuki
4. Genetics:A Conceptual Approach by Benjamin A.Pierce
5. iGenetics: A Molecular Approach by Peter J. Russell
6. Principle of Genetics by Peter Snustad

**UNIT II: ZOO 302.2: Microbiology**

**Course outcome**

*To gain knowledge on importance of microorganisms as primary decomposers, produce food products, produce Antibiotics, synthesize chemicals that our body needs, as test organisms, prevent potential pathogens, Insect Pest Control, Bioremediation, Genetic Engineering. With this exposure, students can be employed in testing/pathology laboratories alternatively.*

**Syllabus:**

1. Outline classification of microorganisms: 5-kingdom, 8-kingdom system, Bergey's manual.
2. Classification & morphology of Bacteria
3. General accounts of Algae, Protists, Fungi & Virus.
4. Microbial Physiology:
  - i. Growth in Bacteria: normal growth curve; methods of measuring growth.
  - ii. Yield and characteristics, strategies of cell division.
  - iii. Bacterial chemotaxis and quorum sensing.
5. Nutrition of microbes
  - i. Principles behind formulating culture media

- ii. Culture techniques; pure cultures.
6. Microbes in soil ecology: fertility, petroleum formation; Role of soil microbial community as a key component of the soil ecosystem.
7. Microbial fermentation: manufacture of industrially important products.

**Reference books:**

- Prescott's Microbiology - Christopher J. Woolverton, Professor, Linda Sherwood, Joanne Willey. 2016.
- Microbiology: An Introduction - Gerard J. Tortora, Berdell R. Funke, Christine L. Case. 2018.

**Course No: ZOO 303A: Fish Taxonomy & Biology and Oceanography**  
(50 Marks, 4 credit)

**Course outcome:**

*A sustainable approach to fisheries and aquaculture will help to protect our natural resources and ensure that fish stocks are available for future generations. Currently, overfishing, ineffective management practices, industrial development and agricultural pollution have reduced fish stocks.*

**Syllabus:**

**UNIT I: ZOO 303A.1: Fish Taxonomy & Biology**

1. Classification of fishes
2. Fish nutrition and growth
3. Fish reproduction and development
4. Fish endocrinology
5. Fish migration

**Reference books / Journal articles:**

1. Helfman, G., B. B. Collette, et al. (2009). The Diversity of Fishes: Biology, Evolution, and Ecology, Wiley. John Wiley & Sons, 2009
2. Nelson, J. S., T. C. Grande, et al. (2016). Fishes of the World, Wiley.Publisher
3. John Wiley & Sons, 2016
4. Jhingran, V. G. (1991). Fish and Fisheries of India, South Asia Books. South Asia Books, 1991

5. Talwar, P. K. and A. G. Jhingran (1991). Inland Fishes of India and Adjacent Countries, Taylor & Francis. CRC Press, 1991

### **UNIT I: ZOO 303A.2: Oceanography**

1. Basic concept of Oceanography
2. Physical oceanography
3. Chemical oceanography
4. Biological oceanography
5. Oceanic resources
6. Oceanic pollution

#### **Reference books / Journal articles:**

1. Trujillo, AP., Thurman, HV. (1983) Essentials of Oceanography. (ISBN-13: 978-0134073545)
2. Garrison, T., Ellis, R. Oceanography: An Invitation to Marine Science. (9<sup>th</sup> Eds).
3. Knauss, JA. Introduction to Physical Oceanography (2<sup>nd</sup> Eds).

### **Course No: ZOO 303B: Biodiversity & Conservation Ecology and Aquatic Ecology**

**(50 Marks, 4 credit)**

#### **Course outcome:**

*Both theory and practical learning process are to acquaint students with both the basic traditional/conventional components of the ecological science in order to develop proper knowledge base to tackle the ongoing ecological changes in and around human settlements with special emphasis to the landscapes & ecosystems of south West Bengal. The entire syllabus has four dimensions- Systems Ecology, Human Ecology, and Aquatic Ecology & Wildlife Ecology. The major emphasis was laid in developing the syllabus to cover not only on traditional aspects of Ecological but also on modern developments in the sphere of ecological sciences: system, mathematical, molecular, urban, restoration and aquatic ecology.*

#### **Syllabus:**

##### **UNIT I: ZOO 303B.1: Biodiversity & Conservation Ecology**

- Biodiversity - Utility and concept. CBD, Megadiversity countries, Biodiversity hotspots. Estimating biodiversity, biodiversity indices. IUCN Red List Category Version 3.1; IUCN



categories of Protected Areas- National Parks, Sanctuaries, Biosphere Reserve. Biodiversity convention, criteria for measuring conservation value of areas. Types of conservation: (i) Ex-situ conservation of animals; captive breeding; species reintroduction, species translocation; population reinforcement; (ii) In-situ conservation- conserving ecosystem function and management. Bioindicators for biodiversity monitoring.

- **Wildlife Ecology: Evolution of Approaches in Wildlife Conservation.** Diversity, ecology, threats & conservation strategies of major Wildlife in West Bengal. Wildlife Habitat management for conservation. Wildlife crime. Social forestry: Joint Forest management- Arabari concept. Biodiversity profile of Ganga River and restoration activities.
- **Conservation of biodiversity– Conservation process; Enhancing & conserving environmental resources.** World Heritage Sites. Red datasheet for India. Critically Endangered Vertebrates of India with special reference to West Bengal. Distribution, habitat utilization, threats to survival of Endangered fauna (Tiger, Wild Elephant), and conservation strategies. Climate change and its effect on wildlife (amphibians & reptiles) and their habitat (eg. alpine vegetation). Human-animal conflict with special reference to elephant migration.
- **Endemic Avifauna of India– IBAs of West Bengal.** Distribution, habitat preference, migration, biology, threats to survival, conservation strategy of Vulture and Great Indian Bustard. Wetland and forest as complementary habitats for the conservation of Avifauna.
- **Tools and techniques for wildlife census and survey.** Technologies for Wildlife Research and Management. Molecular techniques for wildlife biology; DNA fingerprinting in wildlife forensics. Remote sensing: basic idea of GIS and GPS and their application in habitat & wildlife conservation.

#### **Reference books:**

- **Conserving Forest Biodiversity: A Comprehensive Multiscaled Approach-** David B. Lindenmayer, Jerry F. Franklin. 2013.
- **Valuation and Conservation of Biodiversity: Interdisciplinary Perspectives...** Michael Markussen, Ralph Buse, Heiko Garrelts, MaríaManez Costa, Susanne Menzel, Rainer Marggraf. 2005.
- **Practical Approaches to the Conservation of Biological Diversity-** Richard Kenith Baydack, Henry Campa, Jonathan B. Haufler. 1999.

#### **UNIT II: ZOO 303B.2: Aquatic Ecology**

1. **Water as resource** - types and distribution; past changes and present status; Hydrological cycles – different phases, factors contributing to degradation of water quality and management. **Socio-Ecohydrological balancing:** Sustainable water management- surface & groundwater relationships; Base flow, porosity, permeability, transmissivity and storativity.

## 2. Structure and function of aquatic ecosystems and their management : -

- a) Conservation strategies of river, floodplains, lakes, freshwater wetlands, salt marsh and coastal dunes – in respect of climate change.
  - b) **Marine Ecosystem:** Origin, extent and zonation of sea, physical properties and physical processes, chemical composition, behaviour and fate, biological components and their interactions.
  - c) **Coastal Ecosystem:** Definition, extent and types, zonation and geomorphological features, significance, human induced problems, global and marine diversity, integrated coastal zone management.
  - d) **Estuarine Ecosystem:** Definition, classification, structure – biotic assemblage and their interactions, function.
  - e) **Mangrove Ecosystem:** Definition; specialty of this ecosystem; structure and function with special reference to Sundarbans, India; Problems and Management.
  - f) **Coral Ecosystem:** Definition, types and distribution, specialty with regard to biodiversity, productivity and ecosystem functioning, problems and management.
  - g) **Wetland Ecosystem:** Definition, distribution, causal factors, wetland classification, zonation and succession, significance and values, Ramsar sites in India.
  - h) **River Ecosystem:** Fluvial hydrosystem approach; catchment size and drainage basin from selected major rivers, hydrochemical dynamics, biological productivity, human impacts and management perspective.
3. **Wastewater management** – types, source, physical-chemical properties, recycling and bioremediations.
  4. **Aquatic biota, types and trophic interactions** – Macrophytes, phytoplankton, zooplankton, periphyton, benthos and nekton.

### Reference books:

- Limnology: Lake and River Ecosystems - Robert G. Wetzel. 2001.
- Freshwater Ecology: Concepts and Environmental Applications of Limnology - Walter K. Dodds, Matt R Whiles. 2010.

**Course No: ZOO 303C: Genetics and Molecular Biology**

**(50 Marks, 4 credit)**

**Course outcome:**

*The course will acquaint the students to versatile tools and techniques employed in genetic engineering and recombinant DNA technology. A sound knowledge on methodological repertoire allows students to innovatively apply these in basic and applied fields of biological research. Students will introduced to properties, application and limitation of versatile DNA modifying enzymes, gene cloning, sequencing and genetic transformation etc. This course may be deemed as a foundation course serving as a platform for introduction of more advanced cutting-edge technologies that essentially are an amalgamation of basic techniques combined in diverse forms of modern applications. Understand and apply the principles and techniques of molecular biology which prepares students for further education and/or employment in teaching, basic research, or the health professions.*

## **Syllabus:**

### **UNIT I: ZOO 303C.1: Genetics**

#### **1. Organisation of the eukaryotic genome:**

Nucleosome structure, Chromatin remodeling, Histone modification-acetylation, methylation, Centromeric&Telomeric DNA, Epigenetics, Apoptosis.

#### **4. Mechanism of RNA Splicing & Transcription of rRNA gene:**

Production of Mature mRNA in Eukaryotes, 5' and 3' mRNA, RNA editing, transcription of rRNA genes. Self splicing reaction.

#### **2. Mechanism of Crossing Over:**

The holliday model of Crossing over, The Meselson-Radding Model of Recombination, The Double-Strand-Break repair Model of Recombination.

#### **4. Sex determination and Dosage compensation in Mammals and Drosophila:**

Primary and Secondary sex determination in mammals. The Y chromosome sex determinant; Sry, Sox 9 gene product, role of DAX 1 gene, hormonal regulation of the sexual phenotype, Mechanism of sex determination in Drosophila; role of *sxl* gene, transformer gene and double sex protein.

### **Reference Books/ Journal Article:**

1. Introduction to Genetic Analysis by J.F.Griffiths
2. Genes viii by Benjamin Lewin
3. Genetic: Analysis and Principles by Robert J. Brooker
4. An Introduction to the Genetic Analysis by David T. Suzuki

5. Genetics: A Conceptual Approach by Benjamin A. Pierce
6. iGenetics: A Molecular Approach by Peter J. Russell
7. Principle of Genetics by Peter Snustad

## **UNIT II: ZOO 303C.2: Molecular Biology**

### **1. DNA repair mechanism**

Mechanism of DNA mutation, Repair mechanism, Base excision repair, Mismatch Repair, Error prone repair. Recombination repair in *E.coli*

### **2. Signal Transduction pathway**

TGF $\beta$  pathway, Wnt pathway, Jak-Stat pathway, Hedgehog Pathway, Notch Delta, TLR pathway, NF- $\kappa$ B pathway

### **3. Transposable Genetic Element:**

IS element: its property and transposition, Non-composite and Composite Transposition. Cointegration model for transposition. The AC-DC Controlling elements in Corn, Hybrid dysgenesis and P elements in *Drosophila*. Sleeping Beauty model of transposon. Retroposons & retrotransposons

### **4. Genetics of Cancer**

Cell transformation, tumorigenesis, Oncogenes, Tumor suppressor genes, Genomic instability, Epigenetic modification, Angiogenesis, Metastasis and Current therapies.

### **5. Epigenetics and genome imprinting** - DNA methylation in mammals, genomic imprinting in mammals, germ line and pluripotent stem cells, epigenetic control of lymphopoiesis, nuclear transplantation and the reprogramming of the genome. Epigenetics and human disease, epigenetic determinants of cancer.

### **6. Gene Therapy**

Methods of Gene targeting, ex vivo and in- vivo therapy, Stem cell therapy, siRNA and miRNA basics, regulation of transcription and translation of proteins by miRNA. **Transgenesis:** transgenic animal and plants and their application.

### **Reference books:**

1. Molecular Cell Biology by Harvey Lodish
2. Molecular Biology by Robert F. Weaver
3. Molecular Biology of the Gene by James D. Watson
4. iGenetics: A Molecular Approach by Peter J. Russell
5. Human Molecular Genetics by Tom Strachan

6. Introduction to genetics A Molecular Approach by Terry Brown
7. Gene Cloning and Manipulation by Christopher Howe
8. Principles of Gene Manipulation and Genomics by S.B.Primrose

**Course No: ZOO 303D: Diversity & Biology of Parasite and Immunoparasitology**  
(50 Marks, 4 credit)

### **Course outcome**

*The course will acquaint the students to understand host parasite interactions, complement activation, mode of recognition by TLR and its signaling, hypersensitivity with special reference to asthma. The course will enlighten the students regarding different modern diagnostic methods to identify parasitic infections. Vector biology and epidemiology will be another focus to know the propensity of the vector borne diseases and proper formation of vector control.*

### **Syllabus:**

#### **UNIT I: ZOO 303D.1: Diversity and Biology of Parasite**

##### **1. Introduction to Parasitology**

- 1.1 Basic concepts- parasite systematics, ecology and evolution
- 1.2 Host parasite interactions

##### **2. Protozoology**

- 2.1 Classification of Protozoa with suitable example up to subclasses following Levine et al. (1980)
- 2.2 Structure of Apical complex
- 2.3 Life cycle, pathogenicity and control of *Entamoebahistololytica*, *Balantidium coli*
- 2.4 Primary Amoebic Meningoencephalitis

##### **3. Helminthology**

- 3.1 Structure, composition and function of tegument of parasitic helminthes
- 3.2 Life cycle, pathogenicity and control of  
*Diphyllobothriumlatum*, *Echinococcusgranulosus*, *Schistosomahaematobium*,  
*Paragonimuswestermani*, *Trichenellaspiralis*, *Loa loa*
- 3.3 Carbohydrate and protein metabolism in Nematode and Trematode
- 3.4 Different types of scolex in Cestoda
- 3.5 Mode of action of anthelmintic drugs

3.6 Types of cercariae in digenetic trematode

4. Major parasitic diseases of fin fish

## **UNIT II: ZOO 303D.2: Immunoparasitology**

### **1. Immunity and host defense:**

Immune response to the bacteria, Immune response to the Virus, Immune response to the Parasites

### **2. The Complement System**

The Major Pathways of Complement Activation, the Regulation of Complement Activity, Complement Deficiencies

### **3. Hypersensitivity**

Allergy: A Type I Hypersensitivity Reaction, Antibody-Mediated (Type II) Hypersensitivity Reactions, Immune Complex-Mediated (Type III) Hypersensitivity, Delayed-Type (Type IV) Hypersensitivity (DTH), Chronic Inflammation

### **4. Inflammation and Immunity**

Overview of Inflammation, Inflammatory mediators, Inflammation and disease, Therapeutic Aspect

### **5. Transplantation Immunology and Immunomodulation**

Molecular basis of graft vs host reaction. Acute, Hyperacute and chronic Graft rejection, Modern techniques of transplantation (e.g.BMT, liver, cornea etc) Mechanism of action of common immunosuppressant drugs, Immunoboosters

### **6. Tumor Immunology**

Concepts of Immune surveillance, Mechanism of Immune escape by tumor, Anti-tumor Immune response. Modern Immunotherapy of Cancer

### **7. Tolerance and Autoimmunity**

Tolerance and Autoimmunity, Peripheral and Central Tolerance of T and B cell, Malfunction and different autoimmune disease

### **8. Techniques in Clinical parasitology**

Gel electrophoresis, Southern blotting, Western blotting, Immunofluorescence, Immunohistochemistry, Hybridoma technique for MAb, PCR

**Reference books:**

1. Bogitsh, B. J. and Cheng, T. C. (2000). *Human Parasitology*. 2nd Ed. Academic Press, New York.
2. Chandler, A. C. and Read. C. P. (1961). *Introduction to Parasitology*, 10th ed. John Wiley and Sons Inc.
3. Chatterjee, K. D. (1981). *Parasitology (Protozoology and Helminthology)*. 13<sup>th</sup> ed. CBS.
- 4 Cheng, T. C. (1986). *General Parasitology*. 2nd ed. Academic Press, Inc. Orlando. U.S.A.
6. Cox, F. E. G. (1993). *Modern Parasitology*. 2nd ed. Blackwell Scientific Publications. Lea and Febiger, Philadelphia.
- 6 Hati, A. K. (2001). *Medical Parasitology*. Allied Book Agency, Kolkata.
- 7 Noble, E. R. and Noble G. A. (1989). *Parasitology. The Biology of animal Parasites*. 6th ed. Lea and Febiger, Philadelphia.
- 8 Roberts, L. S., Janovy, J. and Nadler S. (2013) *Gerald D. Schmidt & Lary S. Roberts' Foundation of Parasitology*. 9<sup>th</sup> ed. McGraw-Hill International.
- 9 Schmidt, G. D. and Roberts, L. S. (2001). *Foundation of Parasitology*. 3rd ed. McGraw Hill Publishers.
- 10 Schmidt, G. D. (1989). *Essentials of Parasitology*. Wm. C. Brown Publishers (Indian print; 1990, Universal Book Stall).
- 11 .Smyth, J. D. (1994). *Animal Parasitology*. 3rd ed. Cambridge University Press.
12. Soulsby, E. J. L. (1982). *Helminths, Arthropods and Protozoa of domesticated animals*. ELBS and Bailliere Tindall. London.
13. Heinz Mehlhorn (2007) *Parasitology in focus. (Encyclopedic approach)*, 3<sup>rd</sup> Ed. Springer-Verlag, Germany
14. W. Peters and R. Killick-Kendrick (1897) *THE LEISHMANIASIS in Biology and Medicine*. Academic Press (Inc) Ltd.
15. Abbas, A. K., Lichtman, A. H. and Pillai, S. (2006). *Cellular and molecular Immunology*. 6<sup>th</sup> ed. Saunders.
16. Abbas, A. K. and Lichtman, A. H. (2006). *Basic Immunology*. 2<sup>nd</sup> ed. Elsevier.
17. Coico R, Sunshine, G., Benjamini, E. (2003). *Immunology: A short Course*. 5<sup>th</sup> ed. Wiley-Liss: New Jersey.
18. English, L. S. (1994). *Technological Applications of Immunochemicals (BIOTOL)*. Butterworth-Heinemann, Oxford Freeman and Co.
19. Goldsby, R. A., Kindt, T. J., Kuby, J. and Osborne, B. A. (2013). *Immunology*. 7<sup>th</sup> ed. W. H. Freeman and Co.
20. Khan F. H. (2009). *The Elements of Immunology*. Prentice Hall India.
21. Kindt, T., Goldsby, R. Osborne, B. (2007). *Kuby's Immunology*. 6<sup>th</sup> ed. W.H. Freeman and Co.
22. Male, D., Brostoff, J., Roth, D. and Roitt, I. (2006). *Immunology*. 7<sup>th</sup> ed. Mosby.
23. Rao, C. V. (2002). *Immunology*. Narosa Publishing House, New Delhi.
24. Roitt, I. M. and Delves, P. J. (2001). *Roitt's Essential Immunology*. 10<sup>th</sup> ed. Blackwell Science Ltd.

**Course No: C-ZOO 304: Genetics and Haematology (CBCS)**

**(50 Marks, 4 credit)**

**UNIT I: C-ZOO 304.1: Genetics**

**Course outcome**

*The course will provide an understanding of genetic analysis at the gene, genome and population levels. Understanding the Drosophila genetics. Evaluation of the various aspects of structural, functional and comparative genomics. Designing and development of experiments using Drosophila and their evaluation through genetic analysis and interpretations.*

## **Syllabus:**

### **1. Sex-chromosome inheritance**

- i) Chromosomal Determination of Sex, X-linked inheritance
- ii) Pedigree characteristics of Human, X-linked inheritance
- iii) Nondisjunction as proof of the chromosome theory of heredity
- iv) Chromosome theory of Heredity
- v) Sex Determination in *Drosophila*

### **2. Probability in the prediction of progeny distributions**

- i) Using the binomial distribution in genetics
- ii) Testing goodness of fit to a genetic hypothesis, The Chi-Square Method

### **3. Genetic Linkage and Chromosome Mapping**

- i) Linkage and recombination of genes in a chromosome
- ii) The Chi-square test for linkage
- iii) Each pair of linked genes has a characteristics frequency of recombination
- iv) Genetic Mapping, Map distance and frequency of recombination
- v) Crossing –over
- vi) Recombination between genes results from a physical exchange between chromosomes

### **4. Genetic Mapping in a Three-Point Testcross**

- i) Chromosome Interference in a double crossovers
- ii) Genetic Mapping Functions, Genetic Map Distance and physical distance

### **5. Central dogma of Life**

- i) Overview of Organisation of DNA in chromosome
- ii) Basic concept of Replication, transcription and translation

## **Reference books:**

1. Introduction to Genetic Analysis by J.F.Griffiths
2. Genes viii by Benjamin Lewin
3. Genetic: Analysis and Principles by Robert J. Brooker
4. An Introduction to the Genetic Analysis by David T. Suzuki
5. Genetics:A Conceptual Approach by Benjamin A.Pierce
6. iGenetics: A Molecular Approach by Peter J. Russell
7. Principle of Genetics by Peter Snustad



## UNIT II: C-ZOO 304.2: Haematology

### Course outcome

*The course will provide a reflection of physiological homeostasis of human.*

### Syllabus:

1. Haemopoetic tissues: Structure & function [Annelida/ Arthropoda/ Mollusca; Chordata (Class-Fish/mammals)]
2. Blood cells: Structure & function
3. Red cell abnormalities: (Anemia: Introduction, Classification, Iron deficiency anemia & other hypochromic, microcytic anemia)
4. White cells and their disorders: Acute leukemia/ Chronic myeloid leukemia/ Chronic Lymphocytic leukemia
5. Physiology of coagulation & Haemostasis (Congenital bleeding disorders/ Acquired bleeding disorders/ Congenital platelet disorders/Acquired platelet disorders/ Thrombophilic disorders)

## Course No: ZOO 395: Entomology, Ecotoxicology, Molecular Evolution and Microbiology (Practical)

(50 Marks, 4 credit)

### Syllabus:

#### 1) Entomology

- a) Method of collection and preservation of insects
- b) Study of the behavioural modification of legs in honey bee.
- c) Entomological comments on common Pests, Aquatic insects, Insects of medical and economical importance. Galls & Seed cocoon
- d) Mounting of sting apparatus & coupling device of Honey bee.

#### 2) Ecotoxicology

- a) Dose response curve
- b) Lethal dose estimation
- c) MATC in a fish species
- d) Lethal dose 50 mortality curve

#### 3) Molecular Evolution

#### 4) Microbiology

- a. Staining and identification of bacteria, endospores, etc. from a culture media.
- b. Different methods of staining: Gram staining, Negative and differential staining.

- c. Preparation of different culture media with Sterilization techniques.
- d. Inoculation of microbes to respective culture media through proper culture methods.
- e. Enumeration of Coliform bacteria using multiple tube fermentation method.

## **Special paper Practical**

### **Course No: ZOO 396A: Fishery Practical – I & Field trip**

**(50 Marks, 4 credit)**

#### **Syllabus:**

1. Identification of Indian fish fauna
2. Identification of fish food organism/ artificial fish food
3. Dissection – Urinogenital system of Tilapia, ARO of catfishes, Weberianossicles of IMCs.
4. Fecundity estimation
5. Identification of oceanic hemichordates, cephalochordates and urochordates
6. Demonstration of organic carbon, salinity, phosphates and nitrogen
7. Anatomical demonstration of caudal – neural structure of marine fish
8. Field trip

### **Course No: ZOO 396B: Ecology Practical-I & Field trip**

**(50 Marks, 4 credit)**

#### **Syllabus:**

1. Preparation of Climograph
2. Estimation of transparency, TSS, TDS, conductivity, hardness, salinity and alkalinity of water.
3. Estimation of N, P, K content of water/ soil.
4. Basic principle pertaining to acid digestion for the estimation of heavy metals in water sample.
5. Ecological comments on major biotic components in Aquatic system
6. Recording/documentation and submission of terrestrial / aquatic faunal components in and around University campus – (Collection, preservation, identification and analysis of aquatic biota – phytoplankton, zooplankton, benthos, periphyton, aquatic insects, nekton and macrophytes).

7. Applicability of GPS/GIS in recording bioresources and mapping of landscape.
8. Submission of Laboratory notebook.
9. Viva-voce
10. Field trip

**Course No: ZOO 396C: Genetics & Molecular Biology Practical-I & Institute /Lab visit**

**(50 Marks, 4 credit)**

1. Probability in Mendelian Inheritance
  - a. Chi-square, degree of freedom, test for Independence (contingency Chi square),
  - b. Homogeneity Chi-square, Independent Assortment and probability(binomial expansion)
2. Preparation of mitotic metaphase chromosome of rat.
3. Plasmid isolation and restriction digestion and Agarose gel electrophoresis.
4. Lab /Institute visit

**Course No: ZOO 396D: Parasitology Practical-I & Institute /Lab visit**

**(50 Marks, 4 credit)**

**Syllabus:**

1. Stain preparation of blood film from pigeon for identification of protozoan parasite
2. Smear preparation and staining of rectal content of Bufo /gut content of Cockroach for identification of protozoan parasites/examination of gut of fish for identification of endoparasites
3. Whole mount preparation of Ectoparasites
4. Staining and identification of helminth parasites
  - 4.1 Cestode and Trematode with acetocarmine
  - 4.2 Nematode with lactophenol
5. Identification of parasites ( Protozoa, Helminths, Arthropods) with reasons
6. Submission of prepared slides (minimum ten (10) slides)
7. Laboratory Note Book
8. Viva-Voce
9. Lab /Institute visit

# 2<sup>nd</sup> YEAR

## SEMESTER- IV

**Course No: ZOO 401: Environmental Pollution & Management and Biostatistics**  
(50 Marks, 4 credit)

**UNIT I: ZOO 401.1: Environmental Pollution & Management**

### **Course outcome:**

*The students would be provided with current status of environmental pollution and global environmental change. The course would cover biodiversity: status, monitoring and documentation; major drivers of biodiversity change and biodiversity management approaches.*

### **Syllabus:**

1. Global environmental problems; Bioinvasion-Principles, threats and management
2. **Environmental pollution:** Types, natural versus man made; Global scenario.
3. **Air pollution:** Composition of air, zonations of atmosphere; classification, properties/behaviour and fate of air pollutants; properties and role of oxides of nitrogen, and sulphur as air pollutant, green house effect and global warming; photochemical smog, acid rains, effect of pollutants on human health and plants, Noise pollution.
4. **Water pollution:** Classification and behaviour of water pollutants, point and non-point pollution, pollution of water by agricultural wastes (fertilizers and pesticides); sewage, oil, thermal power plants; and eutrophication.
5. **Soil pollution:** Soil pollution through agricultural and solid wastes; soil erosion – types and causative agents; Bioinvasion and its environmental impact; Biosafety and its significance.
6. **Environmental management:** Ecodegradation and pollution; sustainable environmental management; indicators of quality of life. Objectives of conservation; world conservation strategies. Biomonitoring. Green movements; traditional environmental knowledge and people's participation.

### **Reference books:**

1. Biological diversity-Exploiters and Exploited—Paul Hatcher and Nick Battey ( Wiley-Blackwell)

2. The Root Causes of Biodiversity Loss( Edited by Alexander Wood, Pamela Stedman-Edwards, Johanna Mang) EARTHSCAN Publisher
3. The Science of Environmental Pollution—Frank R. Spellman( CRC Press)
4. Environmental Management –N.K.Uberoi9 Excel books)
5. Climate Change-A Multidisciplinary Approach-William James Burroughs( Cambridge University Press)
6. Estuarine and Marine Pollution: Michael J. Kennish( CRC Press)
7. Environmental Biotechnology-A Biosystems Approach-Daniel A. Vallero( Elsevier)
8. Environment, Technology and Development( Edited by Rohan D’Souza0 ( Orient Black Swan)

## UNIT II: ZOO 401.2: Biostatistics

### Course outcome

*To learn about key biostatistical concepts and efficient tools for summarizing and plotting data, make decisions in the presence of uncertainty. Student will be acquainted with parametric and nonparametric statistics, sample and data. The will be able to understand t test, Chi square, correlation, regression and anova.*

### Syllabus:

1. **Concepts of Biostatistics:** Data, population, sample and sampling, frequency distribution, graphical representation of data, parametric and nonparametric statistics
2. **Measures of Central Tendency:**Mean, median and Mode
3. **Measures of Dispersion:** Range, quartile deviation, mean deviation and standard deviation, standard error, variance and covariance
4. **Probability distribution:** Normal distributions, Properties and uses of binomial distributions and Poisson’s distributions
5. **Set theory and probability**
6. **Testing of Hypothesis:** Null Hypothesis. Level of significance. Error of interferenceand degrees of freedom.
7. **Analysis of frequencies:** Chi-square test for goodness of fit.
8. **Student ‘t’ distribution**
9. **Z test and Fisher’s F test**
10. **Correlation and regression:** Properties and types of correlation. Pearson’s product-moment correlation coefficient- properties, assumptions, computation from ungrouped data and

significance test. Partial and multiple correlations. Rank correlation Regressions- types and models, simple linear regression – assumption, properties and computation. Multiple regression.

11. **Analysis of Variances:** Types and models of analysis of variances. Assumption for ANOVA. One-way ANOVA- computation and interpretation of F ratio, multiple comparison t-test, Scheffe's multiple comparison f-test.

12. **Nonparametric Test**

### Reference books:

1. Fundamentals of biostatistics.-7 th ed./Bernard Rosner
2. Principles of Biostatistics/Marcello Pagano/Duxbury Press 1993
3. Statistics in scientific investigation its basis, application and interpretation/Glen McPherson/Springer Verlag 1990
4. Introduction to Biostatistics/Robert R Sokal and F James Rohlf/Dover Publication
5. Biostatistical Analysis, 5th Edition, Jerrold H. Zar
6. Biostatistics by D Das

## Course No: ZOO 402: Developmental Biology and Neuroendocrinology

(50 Marks, 4 credit)

### UNIT I: ZOO 402.1: Developmental Biology

#### Course outcome:

*The course will provide a broad area from embryology to developmental biology. The students will be able to understand the embryonic development, reproductive function and fertilization.*

#### Syllabus:

1. **Early development and molecular mechanism of Amphibian axis formation:**  
Inductive interactions,organisation of a secondary axis,dorsal and ventral signal of the organizer,functions of the organizer, epidermal induction.
2. **Regeneration:**Regeneration of animals with special emphasis on the process of regeneration in Hydra and Amphibia.
3. **Beginning a new organism:** Fertilization in sea urchin and Mammals, the chemoattraction of sperm and egg,species specific binding of acrosomal process, the fast and slow block of polyspermy,role of calcium and egg activation in sea urchin. egg, Translocation and

Capacitation in mammals, recognition at zonapellucida, mouse acrosome reaction and gamete fusion

**Reference books:**

1. Developmental biology, 11th edition 2016 by S. F. Gilbert
2. Principles of Development. Fourth Edition. Lewis Wolpert

**UNIT II: ZOO 402.2: Neuroendocrinology**

**Course outcome**

*The course will provide an understanding of basic concept of neural system. Students would be acquainted with electrical signaling and mechanism. The course will cover evolutionary perspective nervous system, functional organization of the CNS and endocrine disorder*

**Syllabus:**

1. Basic concept of neural system
2. Development and differentiation of neural circuit in vertebrates
3. Neuroendocrine glands in animal
4. Neuro-immuno endocrine pathways
5. Neural/ endocrine disorder

**Reference books:**

1. Brown RE (1994) An Introduction to Neuroendocrinology. Cambridge University Press.
2. Van De Kar, LD. Methods in Neuroendocrinology, C R C Press.
  - Clinical Neuroscience - Kelly Lambert, Craig H. Kinsley. 2004.
  - Text Book of Medical Physiology -Arthur C. Guyton& John Edward Hall. 13<sup>th</sup> Ed.

**Course No: ZOO 403A: Aquaculture & Fish Technology and Inland & Marine fisheries**

**(50 Marks, 4 credit)**

**Course outcome:**

*A sustainable approach to fisheries and aquaculture will help to protect our natural resources and ensure that fish stocks are available for future generations. Currently, overfishing, ineffective management practices, industrial development and agricultural pollution have reduced fish stocks.*

## **Syllabus:**

### **UNIT I: ZOO 403A.1: Aquaculture & Fish Technology**

1. Aquaculture – Problems and prospects in India
2. Integrated fish farming system and fish breeding
3. Fishing methods, harvesting, by-products and export
4. Fish disease
5. Fisheries planning, economics and extension

#### **Reference books:**

1. Stickney, R. R. (2016). Aquaculture, 3rd Edition: An Introductory Text, CABI.
2. Safran, P. (2009). Fisheries and Aquaculture - Volume III. EOLSS Publications, 2009.
3. Pandey, B. N., S. D. Pande, et al. (2007). Aquaculture, A.P.H. Pub.

### **UNIT II: ZOO 403A.2: Inland and Marine fisheries**

1. Freshwater resources/ marine water resources and their biology
2. Trends in aquaculture
3. Estuary
4. Reservoir
5. Waste water management
6. RS-GIS in aquaculture
7. Fishery traits

#### **Reference books:**

1. Jhingran, VG (1975) **Fish and Fisheries of India**. Hindustan Publishing Corporation (India).
2. Nelson, J. (2006) Fishes of the World (4<sup>th</sup> Edition). John Wiley and Sons.
3. Collette, BB, Facey, DE, Helfman, G. (1997) The Diversity of Fishes: Biology, Evolution, and Ecology. Wiley- Blackwell.

### **Course No: ZOO 403B: Systems Ecology & Human Ecology**

**(50 Marks, 4 credit)**

#### **UNIT I: ZOO 403B.1: Systems Ecology**

#### **Course outcome:**



*Both theory and practical learning process are to acquaint students with both the basic traditional/conventional components of the ecological science in order to develop proper knowledge base to tackle the ongoing ecological changes in and around human settlements with special emphasis to the landscapes & ecosystems of south West Bengal. The entire syllabus has four dimensions- Systems Ecology, Human Ecology, and Aquatic Ecology & Wildlife Ecology. The major emphasis was laid in developing the syllabus to cover not only on traditional aspects of Ecological but also on modern developments in the sphere of ecological sciences: system, mathematical, molecular, urban, restoration and aquatic ecology.*

## **Syllabus:**

1. **Community Ecology:** Biotic community: Abundance, Frequency, Relative Abundance, Dominance and Dominance index, Species Diversity and Evenness indices.  $\alpha$ ,  $\beta$ ,  $\gamma$  diversity. Species diversity hypotheses, Species diversity in ecological gradient. Metacommunity concept: Metapopulation structure. Fragmentation of habitat. Metacommunity dynamics: empirical examples.
2. **Restoration Ecology:** Definition, Philosophy and rationale for ecorestoration, Ecological restoration and sustainability, Process of ecorestoration – in the context of landscape to species level.
3. **Ecotourism:** Definition, sustainable development and ecotourism, Foundation of ecotourism, Economics and management issues, merits and demerits.
4. Ecosystem services and human wellbeing. Ecological Economics.
5. **Mathematical Ecology:** Basic concept of ecological modeling; Deterministic and Stochastic models; Theoretical model and analytical solution. - Patterns of Spatial distribution - Random, contagious and regular, coefficient of dispersion. Index of similarity and index of association.
6. **System structure and function:**
  - a. Aquatic system: Physiography of freshwater ecosystems, stratification, distributions and mixing patterns. Dynamics of light, oxygen and nutrient content.
  - b. Terrestrial system: Soils of West Bengal. Ecological processes in Tropical forest ecosystem - Vertical stratification of plants and animals. Production and nutrient cycling. Leaf litter decomposition. Assessment of health of forests / vegetation.

## **Reference books:**

- Elements of Ecology- Thomas Michael Smith, Robert Leo Smith – 2015.
- Ecology: Global Insights and Investigations- Peter Stiling – 2011.

- Issues and Perspectives in Landscape Ecology- John A. Wiens, Michael R. Moss. 2005.
- Handbook of Ecological Restoration – Vol. 2. - Martin R. Perrow, Anthony J. Davy. 2002.

## **UNIT II: ZOO 403B.2: Human Ecology**

- 1. Global Environmental Issues; Global warming** – climate change; Acid rain; Stratospheric ozone layer destruction; Thermal Inversion – Smog, Point and Non-point pollution – fertilizers and pesticides. Carbon sequestration and landscape change.
- 2. Solid waste recycling:** Agriculture, Municipal, Biomedical Wastes – nature, source, environmental impact and management. Wastes in ecosystems and management-urban waste, industrial waste, agricultural waste, radioactive waste, medical waste- effects and control.
- 3. Environmental Management and Acts:** Environmental Impact Assessment: Definition; Types of EIA, EIA process and methodologies – scoping, prediction, evaluation, mitigation and monitoring; Socioeconomic impact assessment; EIA Notification. Environmental Management System, Ecomark.
- 4. Urbanization:** Urban environment – criteria and its present global status, major environmental problems of cities. Urban impact on air and water environment, on biodiversity, agriculture; Indoor Pollution – characteristic of indoor environment, common indoor pollutants, their sources and mode of action; Effect of urbanization on biodiversity.
- 5. Wasteland and watershed management:** Concept – integrated process and mechanism of wasteland restoration and watershed management; Soil erosion – types and factors.
- 6. Bioinvasion:** Related terminologies; Underlying operating principles; Ecological Consequences—Case Studies.

### **Reference books:**

## **Course No: ZOO 403C: Genetic diseases & Molecular Analysis and Applied Genetics**

**(50 Marks, 4 credit)**

### **Course outcome:**

*The course will acquaint the students to versatile tools and techniques employed in genetic engineering and recombinant DNA technology. A sound knowledge on methodological repertoire allows students to innovatively apply these in basic and*

*applied fields of biological research. Students will introduced to properties, application and limitation of versatile DNA modifying enzymes, gene cloning, sequencing and genetic transformation etc. This course may be deemed as a foundation course serving as a platform for introduction of more advanced cutting-edge technologies that essentially are an amalgamation of basic techniques combined in diverse forms of modern applications. Understand and apply the principles and techniques of molecular biology which prepares students for further education and/or employment in teaching, basic research, or the health professions.*

## **Syllabus:**

### **UNIT I: ZOO 403C.1: Genetic Diseases & Molecular Analysis**

#### **1. Molecular & Biochemical basis of**

- a) **Autosomal Disorders:** Cystic fibrosis, Thalassemia
- b) **X-linked Disorders:** Hemophilia A, Muscular dystrophy
- c) **Metabolic Disorders:** Phenylketonuria, Alkaptonuria
- d) **Dynamic Mutations:** Huntington disease
- e) **Late onset disorders:** Alzheimer disease

#### **2. DNA Markers in Genetic Analysis:**

Restriction Fragment Length Polymorphism (RFLP), Tandem Nucleotide Repeat Marker, PCR based markers, Random Amplified Polymorphic DNA (RAPD), Amplification of DNAs by the polymerase Chain reaction and variations, Real time PCR.

#### **3. Protein blotting and Fluorescency:**

Western Blotting techniques for the analysis of Proteins. Mechanism of Fluorescence and Phosphorescence, Fluorescence spectroscopy. In situ localization by FISH.

#### **4. DNA and RNA Sequencing:**

Southern Blotting and Analysis of DNAs, Analysis of RNA by Northern Blot; Sanger Dideoxy Sequencing, Second generation sequencing- pyrosequencing.

#### **5. Recombinant DNA, Plasmid Cloning Vectors:**

Restriction Endonuclease & Methylases, Recombination of DNA fragments, Plasmid cloning vector and expression vectors; Linker DNA, Homopolymer tailing, Blunt end ligation, Shotgun Cloning, cDNA cloning, cDNA microarray, T-A cloning.

**Reference books:**

1. Introduction to Genetic Analysis by J.F.Griffiths
2. Molecular Biology of the Gene by James D. Watson
3. Genes viii by Benjamin Lewin
4. Genetic: Analysis and Principles by Robert J. Brooker
5. Genetics:A Conceptual Approach by Benjamin A.Pierce
6. iGenetics: A Molecular Approach by Peter J. Russell
7. Principle of Genetics by Peter Snustad
8. Concept of Genetics by William S. klug, Michael R. Cummings
9. Human Molecular Genetics by Tom Strachan
10. Introduction to genetics A Molecular Approach by Terry Brown
11. Gene Cloning and Manipulation by Christopher Howe
12. Principles of Gene Manipulation and Genomics by S.B.Primrose

**UNIT II: ZOO 403C.2: Applied Genetics**

1. Genetic changes in differentiation of vertebrate immune cell Genetic control of human antibody, Multigene organization of Ig gene, The mechanism of V(D)J recombination, B cell and T cell receptor genes and expression.
2. The Human Genome Initiative: classical tools of positional cloning, identifying genes mutated in Huntington disease by RFLP, vectors for large scale genome project, studying and comparing Genomic sequences,Human genome.

**Reference books:**

1. Introduction to Genetic Analysis by J.F.Griffiths
2. Molecular Biology of the Gene by James D. Watson
3. Genes viii by Benjamin Lewin
4. Genetic: Analysis and Principles by Robert J. Brooker
5. Genetics:A Conceptual Approach by Benjamin A.Pierce
6. iGenetics: A Molecular Approach by Peter J. Russell
7. Principle of Genetics by Peter Snustad
8. Concept of Genetics by William S. klug, Michael R. Cummings
9. Human Molecular Genetics by Tom Strachan
10. Introduction to genetics A Molecular Approach by Terry Brown
11. Gene Cloning and Manipulation by Christopher Howe
12. Principles of Gene Manipulation and Genomics by S.B.Primrose

**Course No: ZOO 403D: Vector Biology &Vector borne Parasites and  
Molecular Diagnosis & Clinical Parasitology  
(50 Marks, 4 credit)**

**Course outcome:**

*The course will acquaint the students to understand host parasite interactions, complement activation, mode of recognition by TLR and its signaling, hypersensitivity with special reference to asthma. The course will enlighten the students regarding different modern diagnostic methods to identify parasitic infections. Vector biology and epidemiology will be another focus to know the propensity of the vector borne diseases and proper formation of vector control.*

## **Syllabus:**

### **UNIT I: ZOO 403D.1: Vector Biology and Vector borne Parasites**

- 1. General concept about vector- mode of transmission**
- 2. Biology, importance and control of some vectors**
  - 2.1 Mosquito (Anopheles, Culex, Aedes)
  - 2.2 Sand fly
  - 2.3 Fleas
  - 2.4 Ticks
  - 2.5 Mites
  - 2.6 Black fly
  - 2.7 Tsetse fly
- 3. Life cycle, pathogenicity and control of some vector borne parasite**  
*Plasmodium, Leishmania, Wuchereria, Babesia*
- 4. General idea on Zoonosis and Myiasis**
- 5. Lice as vectors of human disease: Typhus, Trench fever, Relapsing fever**
- 6. Dynamics of Filarial transmission**
- 6. Immunology of African Trypanosomiasis**

### **UNIT II: ZOO 403D.2: Molecular Diagnosis & Clinical Parasitology**

1. General concept of molecular diagnosis for parasitic infection
  - 1.1 Advantages and disadvantages of molecular diagnosis
  - 1.2 Fundamental techniques used in molecular diagnosis of endoparasites
2. Biology, clinical and laboratory diagnosis of *Hymenolepis nana*, *Clonorchis sinensis*, *Enterobius vermicularis*, *Dracunculus medinensis*, *Toxoplasma gondii* and *Trichomonas vaginalis*

3. Clinical features of hookworm anaemia
4. Laboratory diagnosis of Amoebiasis
5. Xenodiagnosis of Parasites
6. Parasites as Therapeutic Organisms
7. Marker molecule based immunodiagnosis of endoparasites
  - 7.1 Immunoassay or serological techniques for laboratory diagnosis of endoparasites on the basis of marker molecules like *G.intestinalis*, *B. coli*, *E. histolytica*, *L. donovani*, Malarial parasite using
    - a. ELISA , RIA
    - b. Counter Current Immunoelectrophoresis (CCI)
    - c. Complement Fixation Test (CFT)
    - d.PCR, DNA, RNA probe
8. Epidemiology: Classification, landscape epidemiology, methods of epidemiological studies
  - 8.1 Epidemiology of Malaria, Filariasis, Kala-azar
9. Antigenic variation in Trypanosomiasis: the perfect camouflage

### Reference books:

1. Bogitsh, B. J. and Cheng, T. C. (2000). *Human Parasitology*. 2nd Ed. Academic Press, New York.
2. Chandler, A. C. and Read. C. P. (1961). *Introduction to Parasitology*, 10th ed. John Wiley and Sons Inc.
3. Chandra, G. (2000). *Mosquito*. SreeBhumi Publication Co. Kolkata.
4. Chatterjee, K. D. (1981). *Parasitology (Protozoology and Helminthology)*. 13<sup>th</sup> ed. CBS.
5. Cheng, T. C. (1986). *General Parasitology*. 2nd ed. Academic Press, Inc. Orlando.U.S.A.
6. Cox, F. E. G. (1993). *Modern Parasitology*. 2nd ed. Blackwell Scientific Publications. Lea and Febiger, Philadelphia.
7. Hati, A. K. (2001). *Medical Entomology*. Allied Book Agency, Kolkata.
8. Hati, A. K. (2001). *Medical Parasitology*. Allied Book Agency, Kolkata.
9. Kettle, D. S. (1995). *Medical and veterinary Entomology*. 2<sup>nd</sup> Ed. CAB International.
10. Mullen, G. R. and Durden, L.A. (2009). *Medical and Veterinary Entomology*. 2<sup>nd</sup> Ed. Academic Press.
11. Noble, E. R. and Noble G. A. (1989). *Parasitology. The Biology of animal Parasites*. 6th ed. Lea and Febiger, Philadelphia.
12. Roberts, L. S., Janovy, J. and Nadler S. (2013) *Gerald D. Schmidt & Lary S. Roberts' Foundation of Parasitology*. 9<sup>th</sup> ed. McGraw-Hill International.
13. Schmidt, G. D. and Roberts, L. S. (2001). *Foundation of Parasitology*. 3rd ed. McGraw Hill Publishers.
14. Schmidt, G. D. (1989). *Essentials of Parasitology*. Wm. C. Brown Publishers (Indian print;1990, Universal Book Stall).

15. Smyth, J. D. (1994). *Animal Parasitology*. 3rd ed. Cambridge University Press.
16. Soulsby, E. J. L. (1982). *Helminths, Arthropods and Protozoa of domesticated animals*. ELBS and BailliereTindall. London.
17. R Beaglehole , R Bomta and T Kjelstorm (1993) BASIC EPIDEMIOLOGY Orient Longman in collaboration with WHO , Geneva.
18. W. Peters and R. Killick-Kendrick (1897) THE LEISHMAANIASIS in Biology and Medicine. Academic Press (Inc ) Ltd
19. Lynne Shore Garcia (2010) Dignostic Medical Parasitology 5<sup>th</sup> Edn ASM Press , Washington DC
20. John Hyde ( 1996) Molecular Parasitology Open University Press
21. J Joseph Marr and Miklos Muller (1995) Biochemistry and Molecular Biology of Parasites 2<sup>nd</sup> Edn Academic Press

**Course No: ZOO 494: Environmental Management, Biostatistics, Developmental Biology & Neuro-endocrinology (Practical)**

**(50 Marks, 4 credit)**

**Syllabus:**

1. Biodiversity and Environmental stress
  - i. Qualitative and quantitative estimation of soil and aquatic biodiversity.
  - ii. Basic principles for the estimation of heavy metals.
  - iii. BOD and COD estimation.
  
2. Biostatistics
  - i. Chi square test for goodness of fit with a Mendelian frequency distribution.
  - ii. Computation and significance test of product – moment r between two continuous measurement variables.
  - iii. Computation of simple linear regression.
  - iv. Computation of variance ratio (F) and multiple comparison of Scheffe’s F test for one-way anova and their interpretation.
  - v. Significance of observed sex ratios using binomial distribution.
  
3. Developmentalbiology
  - i. Extraction and identification of different stages of chick embryos (24 hours,48 hours and 72 hours)
  - ii. Histological sectioning and staining of different stages of chick embryo.
  
4. Neuro-Immuno Endocrinology

- i. Neuroendocrine structure demonstration in invertebrates
- ii. Endocrine structure in fish
- iii. Auto-micrograph of neurosecretory elements.
- iv. Neurohistological techniques

**Course No: ZOO 495A: Fishery Practical -II**

**(50 Marks, 4 credit)**

**Syllabus:**

1. Identification of Shellfish, macrophytes and aquatic insects.
2. Physicochemical characteristics of water – salinity, organic carbon, nitrogen, potassium, phosphorus, turbidity and pH.
3. Calculation of - Length weight relationship, gastro-somatic index and gonadosomatic index in IMC.
4. Estimation of muscle protein and lipid from IMC.
5. Identification of freshwater fishes
6. Identification of fish food organism/ aquatic weeds/ aquatic insects
7. Morpho-anatomical demonstration of fish
8. Experimental demonstration of anatomy of fin-fish and shell fish. ARO System of fish.
9. Biochemical parameter demonstration: pH, Dissolve Oxygen, Biological Oxygen Demand, Chemical Oxygen Demand, turbidity, etc.

**Course No: ZOO 495B: Ecology Practical -II**

**(50 Marks, 4 credit)**

**Syllabus:**

1. Estimation of the degree of faunal similarity and association between species.
2. Computation of microdistribution pattern for spatial distribution.
3. Estimation of alpha, beta and gamma diversity.
4. Analysis of the structure of biotic community: Abundance, Relative abundance, Frequency, Species diversity and Dominance indices; Shannon-Weiner diversity index and Importance Value Index.
5. Estimation of textural composition and Water Holding Capacity of soil.



6. Evaluation of Restoration sites; Study of forest/vegetation health- Estimation of tree height, DBH, stand density, canopy density and tree biomass
7. Vermitechnology and related matter: Analysis of biota from urban waste materials & identification of suitable specimen for vermicomposting.
8. Air pollution monitoring: demonstration of Air sampler
9. Submission of Laboratory notebook.
10. Viva-voce

### **Course No: ZOO 495C: Genetics & Mol. Biology Practical -II**

#### **Syllabus:**

1. Probability in Mendelian Inheritance
2. Chi-square, degree of freedom, test for Independence (contingency Chi square),
3. Homogeneity Chi-square, Independent Assortment and probability(binomial expansion)
4. Genetic cross
5. Thin layer chromatography.
6. DNA isolation and Gel electrophoresis (from human blood and goat liver tissue)
7. Family pedigree analysis for autosomal /sex linked, dominant /recessive trait.
8. Isolation & purification of protein & characterization through SDS-PAGE
9. PCR

### **Course No: ZOO 495D: Parasitology Practical-II**

**(50 Marks, 4 credit)**

#### **Syllabus:**

1. Localization of DNA by Fuelgen reaction in protozoa
2. Localization of Glycogen by Periodic Acid Schiff's (PAS) reaction in protozoa
3. Stain preparation and identification of blood parasites from fish
4. Preparation of blood film from human for identification of microfilaria
5. Isolation and electrophoretic separation of plasmid DNA
6. Staining of scolex and proglottids of Cestodes
7. Whole mount preparation of mouth parts of mosquito vector
8. Spot identification of parasites

9. Visit to renowned institute (laboratory)
10. Laboratory Note Book
11. Viva-Voce

## **PROJECT/DISSERTATION**

### **Course outcome:**

*Project report should include introduction, methodology, techniques, results, discussion and bibliography. Institutional and study tour report emphasizing theoretical aspects should be included. Evaluation of the project report and viva-voce will be open defense type through PowerPoint presentation and evaluated by external examiner.*

### **Syllabus:**

<b>Special Paper</b>	<b>Dissertation/ project</b>
<b>ZOO 496A</b>	<b>Fishery</b>
<b>ZOO 496B</b>	<b>Ecology</b>
<b>ZOO 496C</b>	<b>Genetics and Molecular Biology</b>
<b>ZOO 496D</b>	<b>Parasitology</b>